

L9 ANSWER 2 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2008:297521 USPATFULL <<LOGINID::20090129>>  
TITLE: G PROTEIN COUPLED RECEPTORS AND USES THEREOF  
INVENTOR(S): Gaitanaris, George A., Seattle, WA, UNITED STATES  
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Zeng, Hongkui, Shoreline, WA, UNITED STATES  
PATENT ASSIGNEE(S): OMEROS CORPORATION, Seattle, WA, UNITED STATES (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20080260744	A1	20081023
APPLICATION INFO.:	US 2007-940917	A1	20071115 (11)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 2006-527265, filed on 26 Jan 2006, PENDING A 371 of International Ser. No. WO 2003-US28226, filed on 9 Sep 2003		

	NUMBER	DATE
PRIORITY INFORMATION:	US 2002-409303P	20020909 (60)
	US 2003-461329P	20030409 (60)
	US 2006-859469P	20061115 (60)
	US 2006-859473P	20061115 (60)
	US 2006-859470P	20061115 (60)

DOCUMENT TYPE: Utility  
FILE SEGMENT: APPLICATION  
LEGAL REPRESENTATIVE: SEED INTELLECTUAL PROPERTY LAW GROUP PLLC, 701 FIFTH AVE, SUITE 5400, SEATTLE, WA, 98104, US

NUMBER OF CLAIMS: 44  
EXEMPLARY CLAIM: 1-103  
NUMBER OF DRAWINGS: 59 Drawing Page(s)  
LINE COUNT: 16912

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Exemplary diseases and disorders involving the colon include acute self-limited infectious colitis, adenocarcinoma, adenoma, adenoma-carcinoma sequence, adenomatous polyposis coli, adenosquamous carcinomas, allergic (eosinophilic) proctitis and colitis, amebiasis, amyloidosis, angiodysplasia, anorectal malformations, blue rubber bleb nevus syndrome, brown bowel syndrome, Campylobacter fetus infection, carcinoid tumors, carcinoma of the anal canal, carcinoma of the colon and rectum, chlamidial proctitis, Crohn's disease, clear cell carcinomas, Clostridium difficile pseudomembranous enterocolitis, collagenous colitis, colonic adenoma, colonic diverticulosis, colonic inertia, colonic ischemia, congenital atresia, congenital megacolon (Hirschsprung's disease), congenital stenosis, constipation, Cowden's syndrome, cystic fibrosis, cytomegalovirus colitis, diarrhea, dieulafor lesion, diversion colitis, diverticulitis, diverticulosis, drug-induced diseases, dysplasia and malignancy in inflammatory bowel disease, Ehlers-Danlos syndromes, enterobiasis, familial adenomatous polyposis, familial polyposis syndromes, Gardner's syndrome, gastrointestinal stromal neoplasms, hemangiomas and vascular anomalies, hemorrhoids, hereditary hemorrhagic telangiectasia, herpes colitis, hyperplastic polyps, idiopathic inflammatory bowel disease, incontinence,

inflammatory bowel syndrome, inflammatory polyps, inherited adenomatous polyposis syndromes, intestinal hamartomas, intestinal pseudo-obstruction, irritable bowel syndrome, ischemic colitis, juvenile polyposis, juvenile polyps, Klippel-Trenaunay-Weber syndrome, leiomyomas, lipomas, lymphocytic (microscopic) colitis, lymphoid hyperplasia and lymphoma, malaknock outplakia, malignant lymphoma, malignant neoplasms, malrotation, metastatic neoplasms, mixed hyperplastic and adenomatous polyps, mucosal prolapse syndrome, neonatal necrotizing enterocolitis, neuroendocrine cell tumors, neurogenic tumors, neutropenic enterocolitis, non-neoplastic polyps, Peutz-Jeghers syndrome, pneumatosis cystoides intestinalis, polyposis coli, pseudomembranous colitis, pseudoxanthoma elasticum, pure squamous carcinomas, radiation colitis, schistosomiasis, Shigella colitis ( bacilliary dysentery), spindle cell carcinomas, spirochetosis, stercolar ulcers, stromal tumors, systemic sclerosis and CREST syndrome, trichuriasis, tubular adenoma (adenomatous polyp, polypoid adenoma), Turcot's syndrome, Turner's syndrome, ulcerative colitis, villous adenoma, and volvulus.

DETD The results of RT-PCR analysis with 100 different GPCRs and 26 mouse tissues (17 peripheral tissues and 9 brain regions) are shown in FIG. 4. The data is presented as a semi-quantitative scattergram. The most remarkable finding was that 94% of GPCRs were detected in the brain, generally in 4 to 5 distinct anatomical areas. The largest number of genes was detected in the hypothalamus (82 genes), a brain region of high structural complexity. Individual peripheral tissues also showed expression of multiple different GPCRs, ranging from 12 genes in muscle to 69 genes in ovary.

L9 ANSWER 3 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2008:297514 USPATFULL <<LOGINID::20090129>>  
 TITLE: COMBINATION OF BLYS AND/OR APRIL INHIBITION AND IMMUNOSUPPRESSANTS FOR TREATMENT OF AUTOIMMUNE DISEASE  
 INVENTOR(S): Ponce, Rafael A., Seattle, WA, UNITED STATES  
 Wallis, Wayne J., Seattle, WA, UNITED STATES  
 Holdren, Matthew S., Seattle, WA, UNITED STATES  
 Zuckerman, Linda, Seattle, WA, UNITED STATES  
 Littau, Alisa M., Woodinville, WA, UNITED STATES  
 Van Ness, Kirk P., Bainbridge Island, WA, UNITED STATES  
 Pena Rossi, Claudia, Geneva, SWITZERLAND  
 Graffner, Hans Otto Lennart, Helsingborg, SWEDEN

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20080260737	A1	20081023
APPLICATION INFO.:	US 2008-57133	A1	20080327 (12)

	NUMBER	DATE
PRIORITY INFORMATION:	US 2007-908365P	20070327 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	ZYMOGENETICS, INC., INTELLECTUAL PROPERTY DEPARTMENT, 1201 EASTLAKE AVENUE EAST, SEATTLE, WA, 98102-3702, US	
NUMBER OF CLAIMS:	44	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	3 Drawing Page(s)	
LINE COUNT:	4296	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes for this purpose include but are not limited to eubacteria,

such as Gram-negative or Gram-positive organisms, for-example, Enterobacteriaceae such as Escherichia, e.g., E. coli, Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. licheniformis (e.g., B. licheniformis 41P disclosed in DD 266,710 published 12 Apr. 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. Preferably, the host cell should secrete minimal amounts of proteolytic enzymes.

DETD The percent of CD3-CD40+B cells of total lymphocytes in samples can be obtained by the following gating strategy. The lymphocyte population is marked on the forward scatter/side scattergram to define Region 1(Ri). Using events in RI, fluorescence intensity dot plots are displayed for CD40 and CD3 markers. Fluorescently labeled isotype controls are used to determine respective cutoff points for CD40 and CD3 positivity.

L9 ANSWER 4 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2008:207495 USPATFULL <<LOGINID::20090129>>  
 TITLE: Polypeptides That Bind Baff And/Or April  
 INVENTOR(S): Kelley, Robert F., San Bruno, CA, UNITED STATES  
 PATENT ASSIGNEE(S): Genentech, Inc., South San Francisco, CA, UNITED STATES (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20080181886	A1	20080731
APPLICATION INFO.:	US 2005-666781	A1	20051028 (11)
	WO 2005-US39154		20051028
			20070910 PCT 371 date

	NUMBER	DATE
PRIORITY INFORMATION:	US 2004-625341P	20041104 (60)
	US 2005-673127P	20050419 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	CLARK & ELBING LLP, 101 FEDERAL STREET, BOSTON, MA, 02110, US	
NUMBER OF CLAIMS:	81	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	6 Drawing Page(s)	
LINE COUNT:	4761	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes for this purpose include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as Escherichia, e.g., E. coli, Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. lichenifonnis (e.g., B. lichenifonnis 41P disclosed in DD 266,710 published 12 Apr. 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. Preferably, the host cell should secrete minimal amounts of proteolytic enzymes.

DETD Peripheral B-cell concentrations are determined by a FACS method that count CD3-/CD40+ cells. The percent of CD3-CD40+B cells of total lymphocytes in samples can be obtained by the following gating strategy. The lymphocyte population is marked on the forward scatter/side scattergram to define Region 1 (R1). Using events in R1, fluorescence intensity dot plots are displayed for CD40 and CD3 markers. Fluorescently labeled isotype controls are used to determine respective

cutoff points for CD40 and CD3 positivity.

L9 ANSWER 5 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2008:183415 USPATFULL <<LOGINID::20090129>>  
TITLE: METHOD FOR ACTIVATING AN ANTIGEN, METHOD FOR DETECTING  
A CELL, AND SOLUTION FOR ACTIVATING AN ANTIGEN  
INVENTOR(S): YASUDA, Yuichi, Kobe-shi, JAPAN  
Morita, Masakatsu, Kobe-shi, JAPAN  
Ding, Junyi, Kobe-shi, JAPAN  
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Kishi, Kazuki, Kobe-shi, JAPAN  
PATENT ASSIGNEE(S): SYSMEX CORPORATION, Kobe-shi, JAPAN (non-U.S.  
corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20080160542	A1	20080703
APPLICATION INFO.:	US 2007-966318	A1	20071228 (11)

	NUMBER	DATE
PRIORITY INFORMATION:	JP 2006-355952	20061228
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	SUGHRUE MION, PLLC, 2100 PENNSYLVANIA AVENUE, N.W., SUITE 800, WASHINGTON, DC, 20037, US	
NUMBER OF CLAIMS:	20	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	7 Drawing Page(s)	
LINE COUNT:	773	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DRWD FIG. 3 (A) shows the scattergram of unheated cells, and FIG. 3 (B) shows a scattergram of heated cells.

DRWD FIG. 4-1 shows the scattergram of cells of proliferative phase obtained from uterine cervix, and FIG. 4-2 shows the scattergram of cells of the secretory phase obtained from uterine cervix.

DETD Cervical cancer is diagnosed using cells extracted from a cervix. It is known that cervical cells remarkably change in their cell morphology depending on the stage in the menstrual cycle. More specifically, relatively strong cells with stable morphology are abundant in the early, middle, and later proliferative phases when estrogen is released. In the early, middle, and later secretory phases when progesterone is released, Doderlein's bacillus appears and dissolves the cells, so that bare nuclei appear, and impurities such as erythrocytes and mucus increase.

DETD Diagnosis of cervical cancer requires squamous cells. The cells keep their shapes to a degree in the proliferative phase with no breakage by swelling or other causes, so that their images appear in the region expressed by the chained line (a) in FIG. 3A (squamous cell-appearing region). On the other hand, images of impurities such as bare nuclei and broken cells appear in the region expressed by the dashed line (b) in FIG. 3A (impurity-appearing region). For example, when the specimen used in FIG. 3A is heated at 100° C. under the conventional antigen activation method, as shown in FIG. 3B, bare nuclei and cell fragments increase, and less cells keep their shapes. In addition, the specimen in the secretory phase contains much cells dissolved by the influence of Doderlein's bacillus, so that lots of bare nuclei and dissolved cells appear in the lower region of a graph as expressed by the region (b) in FIG. 3B (impurity-appearing region). The horizontal axis of FIG. 3 represents cell circularity (a cell approaches a circle or round as the value of circularity approaches to the left end, and

increases in asperities or irregularity as the value approached to the right end), and the vertical axis represents the cell area.

DETD The same specimens in the early, middle, and later proliferative phases, and early, middle, and later secretory phases as those used in Example 2 were individually subjected to the antigen activation treatment with an antigen activation solution containing 15 w/v % of urea in the same manner as Example 2. After the treatment, the specimen was subjected to tyramide staining, and the specimen containing the stained cells was dropped onto a glass slide. The glass slide was mounted on an inverted microscope, AxioVert 200 manufactured by Zeiss (condenser: LD condenser (N.A.0.55) Ph2, objective lens: 20 times, LD AchroPlan (N.A.0.4) Ph2, fluorescent filter: filter set #17), and the cells on the glass slide were imaged with an exposure time of 1 second using a CCD camera, AxioCamHRC manufactured by Zeiss. The image was analyzed by Image-Pro Plus (ver. 4.5.1.23) manufactured by Media Cybernetics, and the area and circularity of the imaged cells were calculated. On the basis of the calculation result, a scattergram composed of two axes of cell area (vertical axis) and circularity (horizontal axis) was prepared.

DETD The scattergram is shown in FIG. 4. As is evident from FIG. 4, for all the specimens in any stages of the menstrual cycle, the number of cells appearing in the squamous cell appearing region scarcely changed regardless the antigen activation treatment with the antigen activation solution. This fact indicate that squamous cells essential for the diagnosis of cervical cancer little change in their morphology.

L9 ANSWER 6 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2008:73615 USPATFULL <<LOGINID::20090129>>

TITLE: Hematopoietic growth factor inducible neurokinin-1 gene and uses thereof

INVENTOR(S): Rameshwar, Pranela, Maplewood, NJ, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20080064649	A1	20080313
APPLICATION INFO.:	US 2007-782185	A1	20070724 (11)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 2003-463106, filed on 17 Jun 2003, PENDING Continuation-in-part of Ser. No. US 2001-39272, filed on 20 Oct 2001, GRANTED, Pat. No. US 6939955		

	NUMBER	DATE
PRIORITY INFORMATION:	US 2000-241881P	20001020 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	Licata & Tyrrell P.C., 66 E. Main Street, Marlton, NJ, 08053, US	
NUMBER OF CLAIMS:	12	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	7 Drawing Page(s)	
LINE COUNT:	3495	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Representative examples of appropriate hosts for in vitro procedures include bacterial cells, such as streptococci, staphylococci, E. coli, Streptomyces and Bacillus subtilis cells; fungal cells, such as yeast cells and Aspergillus cells, insect cells such as Drosophila S2 and Spodoptera Sf9 cells; animal cells such as CHO, COS, HeLa, C127, 3T3, BHK, HEK 293 and Bowes melanoma cells, and plant cells. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein.

DETD Another experiment of the present invention characterizes slow-growing

and/or drug resistant clones by flow cytometry, which determines the degree that cells from different clones can pump dye (either Rhodamine 123 or Hoechst as used experimentally). The cancer stem cells are likely more efficient than cancer progenitors to pump dye out of cells. The size and scatter pattern of the different clones are examined to determine whether the slow-growing clones represent side population (S-Pop) cells and whether the progenitor cells larger so that they would be identified at a particular region in the scattergram. A subset of the study population is collected by cell sorting based on size and/or rhodamine uptake. Drug resistant cells are categorized as S-Pop, S-Pop/Rhodamine or Hoescht.sup.dim, S-Pop/Rhodamine or Hoescht.sup.bright, Forward scatter (FSc), FSc/Rhodamine or Hoescht.sup.dim; FSc/Rhodamine or Hoesch.sup.bright Cancer cells are subsequently stimulated in a third round of selection, which is significant because it assists in understanding how a cancer stem cell could convert into an aggressive phenotype and form progenitors that metastasize to tertiary sites. Clones that have been narrowed as potential cancer stem cells are used. Cells are always re-cultured with the anti-cancer agents prior to assays so as to be certain that the experiments are performed with clones that are resistant to the high concentration of drugs. Cells are then studied to determine if they can be stimulated to self-renew and also form cancer progenitors.

L9 ANSWER 7 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2008:43640 USPATFULL <<LOGINID::20090129>>  
 TITLE: ILT3 binding molecules and uses therefor  
 INVENTOR(S): Ponath, Paul, San Francisco, CA, UNITED STATES  
 Rosenzweig, Michael, Boston, MA, UNITED STATES  
 Ponte, Jose F., South Boston, MA, UNITED STATES  
 PATENT ASSIGNEE(S): TolerRX, Inc., Cambridge, MA, UNITED STATES (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20080038260	A1	20080214
APPLICATION INFO.:	US 2007-820363	A1	20070619 (11)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 2006-471397, filed on 19 Jun 2006, PENDING		

	NUMBER	DATE
PRIORITY INFORMATION:	US 2006-814931P	20060619 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	LAHIVE & COCKFIELD, LLP, ONE POST OFFICE SQUARE, BOSTON, MA, 02109-2127, US	
NUMBER OF CLAIMS:	43	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	6 Drawing Page(s)	
LINE COUNT:	4909	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD E. coli is one prokaryotic host particularly useful for cloning the polynucleotides (e.g., DNA sequences) of the present invention. Other microbial hosts suitable for use include bacilli, such as Bacillus subtilus, and other enterobacteriaceae, such as Salmonella, Serratia, and various Pseudomonas species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (trp) promoter system, a beta-lactamase

promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing transcription and translation.

DETD Typical antigens of interest may be classified as follows: protein antigens, such as ceruloplasmin and serum albumin; bacterial antigens, such as teichoic acids, flagellar antigens, capsular polysaccharides, and extra-cellular bacterial products and toxins; glycoproteins and glycolipids; viruses, such as animal, plant, and bacterial viruses; conjugated and synthetic antigens, such as proteinhaptens conjugates, molecules expressed preferentially by tumors, compared to normal tissue; synthetic polypeptides; and nucleic acids, such as ribonucleic acid and deoxyribonucleic acid. The term "infectious agent," as used herein, includes any agent which expresses an antigen which elicits a host cellular immune response. Non-limiting examples of viral antigens which may be considered useful as include, but are not limited to, the nucleoprotein (NP) of influenza virus and the Gag proteins of HIV. Other heterologous antigens include, but are not limited to, HIV Env protein or its component parts gp120 and gp41, HIV Nef protein, and the HIV Pol proteins, reverse transcriptase and protease. In addition, other viral antigens such as Ebola virus (EBOV) antigens, such as, for example, EBOV NP or glycoprotein (GP), either full-length or GP deleted in the mucin region of the molecule (Yang Z-Y, et al. (2000) Nat Med 6:886-9, 2000), small pox antigens, hepatitis A, B or C virus, human rhinovirus such as type 2 or type 14, Herpes simplex virus, poliovirus type 2 or 3, foot-and-mouth disease virus (FMDV), rabies virus, rotavirus, influenza virus, coxsackie virus, human papilloma virus (HPV), for example the type 16 papilloma virus, the E7 protein thereof, and fragments containing the E7 protein or its epitopes; and simian immunodeficiency virus (SIV) may be used. The antigens of interest need not be limited to antigens of viral origin. Parasitic antigens, such as, for example, malarial antigens are included, as are fungal antigens, bacterial antigens and tumor antigens. Examples of antigens derived from bacteria are those derived from *Bordetella pertussis* (e.g., P69 protein and filamentous haemagglutinin (FHA) antigens), *Vibrio cholerae*, *Bacillus anthracis*, and *E. coli* antigens such as *E. coli* heat labile toxin B subunit (LT-B), *E. coli* K88 antigens, and enterotoxigenic *E. coli* antigens. Other examples of antigens include *Schistosoma mansoni* P28 glutathione S-transferase antigens (P28 antigens) and antigens of flukes, mycoplasma, roundworms, tapeworms, *Chlamydia trachomatis*, and malaria parasites, e.g., parasites of the genus *plasmodium* or *babesia*, for example *Plasmodium falciparum*, and peptides encoding immunogenic epitopes from the aforementioned antigens.

DETD An infection, disease or disorder which may be treated or prevented by the administration of a composition of the invention includes any infection, disease or disorder wherein a host immune response acts to prevent the infection, disease or disorder. Diseases, disorders, or infection which may be treated or prevented by the administration of the compositions of the invention include, but are not limited to, any infection, disease or disorder caused by or related to a fungus, parasite, virus, or bacteria, diseases, disorders or infections caused by or related to various agents used in bioterrorism, listeriosis, Ebola virus, SARS, small pox, hepatitis A, hepatitis B, hepatitis C, diseases and disorders caused by human rhinovirus, HIV and AIDS, Herpes, polio, foot-and-mouth disease, rabies, diseases or disorders caused by or related to: rotavirus, influenza, coxsackie virus, human papilloma virus, SIV, malaria, cancer, e.g., tumors, and diseases or disorders caused by or related to infection by *Bordetella pertussis*, *Vibrio cholerae*, *Bacillus anthracis*, *E. coli*, flukes, mycoplasma, roundworms, tapeworms, *Chlamydia trachomatis*, and malaria parasites, etc.

DETD As used herein, the term "bacterial infections" include infections with a variety of bacterial organisms, including gram-positive and gram-negative bacteria. Examples include, but are not limited to, *Neisseria* spp, including *N. gonorrhea* and *N. meningitidis*, *Streptococcus* spp, including *S. pneumoniae*, *S. pyogenes*, *S. agalactiae*, *S. mutans*; *Haemophilus* spp, including *H. influenzae* type B, non typeable *H. influenzae*, *H. ducreyi*; *Moraxella* spp, including *M. catarrhalis*, also known as *Branhamella catarrhalis*; *Bordetella* spp, including *B. pertussis*, *B. parapertussis* and *B. bronchiseptica*; *Mycobacterium* spp., including *M. tuberculosis*, *M. bovis*, *M. leprae*, *M. avium*, *M. paratuberculosis*, *M. smegmatis*; *Legionella* spp, including *L. pneumophila*; *Escherichia* spp, including enterotoxigenic *E. coli*, enterohemorrhagic *E. coli*, enteropathogenic *E. coli*; *Vibrio* spp, including *V. cholera*, *Shigella* spp, including *S. sonnei*, *S. dysenteriae*, *S. flexnerii*; *Yersinia* spp, including *Y. enterocolitica*, *Y. pestis*, *Y. pseudotuberculosis*, *Campylobacter* spp, including *C. jejuni* and *C. coli*; *Salmonella* spp, including *S. typhi*, *S. paratyphi*, *S. choleraesuis*, *S. enteritidis*; *Listeria* spp., including *L. monocytogenes*; *Helicobacter* spp, including *H. pylori*; *Pseudomonas* spp, including *P. aeruginosa*, *Staphylococcus* spp., including *S. aureus*, *S. epidermidis*; *Enterococcus* spp., including *E. faecalis*, *E. faecium*; *Clostridium* spp., including *C. tetani*, *C. botulinum*, *C. difficile*; *Bacillus* spp., including *B. anthracis*; *Corynebacterium* spp., including *C. diphtheriae*; *Borrelia* spp., including *B. burgdorferi*, *B. garinii*, *B. afzelii*, *B. andersonii*, *B. hermsii*; *Ehrlichia* spp., including *E. equi* and the agent of the Human Granulocytic Ehrlichiosis; *Rickettsia* spp, including *R. rickettsii*; *Chlamydia* spp., including *C. trachomatis*, *C. pneumoniae*, *C. psittaci*; *Leptospira* spp., including *L. interrogans*; *Treponema* spp., including *T. pallidum*, *T. denticola*, *T. hyodysenteriae*. Preferred bacteria include, but are not limited to, *Listeria*, *mycobacteria*, *mycobacteria* (e.g., *tuberculosis*), *Anthrax*, *Salmonella* and *Listeria monocytogenes*.

DETD Intracellular calcium flux studies using flow cytometry analysis was performed as described by Rabin, et al. (J Immunol. (1999)162:3840-3850). Briefly, monocyte-derived dendritic cells (2+10<sup>sup.7</sup>) were suspended in HBSS-HEPES (HBSS supplemented with 10 mM HEPES, C<sup>sup.a++</sup>, Mg<sup>sup.++</sup>, and 1% fetal calf serum). Indo-1 and pleuronic detergent (Molecular Probes, Eugene, Oreg.) were added at final concentrations of 5  $\mu$ M and 300  $\mu$ g/mL, respectively. The cell suspension was incubated at 30° C. for 45 minutes with gentle agitation. Cells were then washed twice with the HBSS-HEPES, stained with anti-CD1a<sup>sup.+</sup>, and washed again. Calcium flux for CD1a<sup>sup.+</sup>dendritic cells was performed using a FACSVantage flow cytometer (Becton Dickinson) equipped with an argon laser tuned to 488 nM and a krypton laser tuned to 360 nM. Indo-1 fluorescence was analyzed at 390/20 nM and 530/20 nM for bound and free calcium, respectively. Before stimulation, cell suspensions were warmed at 37° C. for 3 minutes. The CD1a<sup>sup.+</sup> cell population was gated, and baseline fluorescent ratios were collected for 30 seconds. Cells were then stimulated with either fMLP (10<sup>sup.5</sup> M), T-20 peptide (10<sup>sup.5</sup> M), or F-peptide (10<sup>sup.5</sup> M) followed by fMLP (10<sup>sup.8</sup> M). Collections continued until calcium flux returned to basal levels. Changes in Indo-1 fluorescence were expressed as the ratio of bound to free intracellular calcium, and scattergrams represented the entire CD1a<sup>sup.+</sup> cell population at the time of stimulation. Data analysis was performed using Flowjo software (Tree Star, San Carlos, Calif.).

L9 ANSWER 8 OF 42 CAPLUS COPYRIGHT 2009 ACS on STN

ACCESSION NUMBER: 2008:1343141 CAPLUS <<LOGINID::20090129>>

DOCUMENT NUMBER: 149:528543

TITLE: Proposed MIC and disk diffusion microbiological cutoffs and spectrum of activity of retapamulin, a



AUTHOR(S): novel topical antimicrobial agent  
 Traczewski, Maria M.; Brown, Steven D.  
 CORPORATE SOURCE: The Clinical Microbiology Institute, Inc.,  
 Wilsonville, OR, USA  
 SOURCE: Antimicrobial Agents and Chemotherapy (2008), 52(11),  
 3863-3867  
 CODEN: AMACCQ; ISSN: 0066-4804  
 PUBLISHER: American Society for Microbiology  
 DOCUMENT TYPE: Journal  
 LANGUAGE: English  
 REFERENCE COUNT: 23 THERE ARE 23 CITED REFERENCES AVAILABLE FOR THIS  
 RECORD. ALL CITATIONS AVAILABLE IN THE RE FORMAT

AB Retapamulin, the first pleuromutilin antimicrobial agent approved for the  
 topical treatment of skin infections in humans, was tested against 987  
 clin. isolates representing 30 species and/or resistance groups. MICs  
 were determined along with disk diffusion zone diams. using a 2- $\mu$ g disk.  
 Population distribution and MIC vs. disk zone diameter scattergrams  
 were analyzed to determine microbiol. MIC cutoff values and inhibition zone  
 correlates. Min. bactericidal concns. were performed on a smaller subset  
 of key species. The retapamulin MIC90 against 234 Staphylococcus aureus  
 isolates and 110 coagulase-neg. staphylococci was 0.12  $\mu$ g/mL.  
 Retapamulin MIC90s ranged from 0.03 to 0.06  $\mu$ g/mL against  
 beta-hemolytic streptococci including 102 Streptococcus pyogenes, 103  
 Streptococcus agalactiae, 59 group C Streptococcus, and 71 group G  
 Streptococcus isolates. The MIC90 against 55 viridans group streptococci  
 was 0.25  $\mu$ g/mL. Retapamulin had very little activity against 151  
 gram-neg. bacilli and most of the Enterococcus species tested.  
 Based on the data from this study, for staphylococci, MICs of  $\leq 0.5$ ,  
 1, and  $\geq 2$   $\mu$ g/mL with corresponding disk diffusion values of  
 $\geq 20$  mm, 17 to 19 mm, and  $\leq 16$  mm can be proposed for  
 susceptible, intermediate, and resistant microbiol. cutoffs, resp. For  
 beta-hemolytic streptococci, a susceptible-only MIC of  $\leq 0.25$   
 $\mu$ g/mL with a corresponding disk diffusion value of  $\geq 15$  mm can be  
 proposed for susceptible-only microbiol. cutoffs.

IT Bacilli  
 (gram-neg.; proposed MIC and disk diffusion microbiol. cutoffs and  
 spectrum of activity of retapamulin as a novel topical antimicrobial  
 agent)

L9 ANSWER 9 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2007:308798 USPATFULL <<LOGINID::20090129>>  
 TITLE: Apparatus for analyzing particles in urine and method  
 thereof  
 INVENTOR(S): Tanaka, Yousuke, Kobe, JAPAN  
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 PATENT ASSIGNEE(S): SYSMEX CORPORATION, Hyogo, JAPAN (non-U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20070269897	A1	20071122
APPLICATION INFO.:	US 2007-798113	A1	20070510 (11)

	NUMBER	DATE
PRIORITY INFORMATION:	JP 2006-138557	20060518
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	SUGHRUE MION, PLLC, 2100 PENNSYLVANIA AVENUE, N.W., SUITE 800, WASHINGTON, DC, 20037, US	

NUMBER OF CLAIMS: 20  
EXEMPLARY CLAIM: 1  
NUMBER OF DRAWINGS: 13 Drawing Page(s)  
LINE COUNT: 1028  
CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM Leukocytes are frequently found in urine samples from patients with renal infection, urinary tract infection, renal tuberculosis or the like. Therefore, it is possible to detect inflammation and infection at earlier stage through measurements of leukocytes in the urine sample. Leukocytes are from about 6 to 14  $\mu\text{m}$  in size. Measurement of bacteria is an examination to check presence or absence of infection. The bacteria include cocci and bacilli. Cocci are spherical bacteria from about 0.5 to 2  $\mu\text{m}$  in size, while bacilli are bacteria having a major axis in the range of about 2 to 10  $\mu\text{m}$ . Cocci, if proliferated, result in a conglomeration of a chained shape representing an in-line moniliform or of a grape shape representing an irregularly and botryoidally-aggregated ones.

DRWD FIGS. 11(a) to 11(e) are drawings showing one example of a scattergram obtained by the apparatus for analyzing particles in urine relating to one embodiment according to the present invention;

DRWD FIG. 12 is a drawing showing one example of a scattergram of a bacteria system obtained by the apparatus for analyzing particles in urine relating to one embodiment according to the present invention; and

DRWD FIG. 13 is a drawing showing one example of a scattergram of a bacteria system obtained by the apparatus for analyzing particles in urine relating to one embodiment according to the present invention.

DETD Then, raw data of the urinary particles (SED) are generated in the personal computer 13 (Step S28) and at the same time, a scattergram is generated based on the data (Step S29). Then, clustering of the scattergram prepared by algorithm analysis is performed (Step S30), and the number of particles is counted for every cluster (Step S31).

DETD Then, they are transmitted to the personal computer 13 via the LAN adapter 12. Raw data of the bacteria (BAC) are generated in the personal computer 13 (Step S35), and a scattergram is generated based on the data (Step S36). Then, clustering of the scattergram prepared as mentioned by algorithm analysis is performed (Step S37), and the number of particles is counted for every cluster (Step S38). Results of the measurement obtained as mentioned above are displayed on a display which is a display means of the personal computer 13 (Step S39).

DETD As measurement results of the urinary particles (SED), scattergrams are generated from each of signals of forward-scattered light, side-scattered light, and fluorescence. FIG. 11(a) is a scattergram in which the horizontal axis represents fluorescence intensity (low-sensitivity) (FLL) and the vertical axis represents forward-scattered light intensity (FSC). Epithelial cells (EC) and leukocytes (WBC), which are large cells having nuclei, appear in a region of strong fluorescence signal intensity. Majority of epithelial cells are larger in cell size than leukocytes and appear in a region where fluorescence intensity is stronger than that of leukocytes, while the range of appearance of some small-sized epithelial cells overlaps with that of leukocytes. In order to distinguish the both, a side-scattered light signal is used. FIG. 11(b) is a scattergram in which the horizontal axis represents side-scattered light intensity (SSC) and the vertical axis represents forward-scattered light intensity (FSC). Since epithelial cells appear in a region where side-scattered light intensity is stronger than leukocytes, epithelial cells are identified from this scattergram.

DETD FIG. 11(c) is a scattergram in which the horizontal axis

represents fluorescence intensity (high-sensitivity) (FLH) and the vertical axis represents forward-scattered light intensity (FSC) and shows a region where fluorescence intensity is low. Erythrocytes (RBC) have no nuclei and therefore are found in regions where fluorescence intensity is low. Some crystals (X'TAL) appear in regions of erythrocytes appearance, and therefore, a side-scattered light signal is used for confirmation of appearance of crystals. FIG. 11(b) is a scattergram in which the horizontal axis represents side-scattered light intensity (SSC) and the vertical axis represents forward-scattered light intensity (FSC). With crystals, the center of distribution of side-scattered light intensity is not constant, crystals appear in regions where the intensity is high, and therefore, discrimination from erythrocytes is performed from this scattergram.

DETD FIG. 11(d) is a scattergram in which the horizontal axis represents fluorescence width (FLLW) and the vertical axis represents fluorescence width 2 (FLLW2). FLLW indicates a width of a fluorescence signal to capture particles in which cell membranes are stained and FLLW2 indicates a width of a stronger fluorescence signal such as nuclei. As shown in the drawing, FLLW of casts (CAST) is greater and FLLW2 of casts with contents (P. CAST) is greater. Further, casts without contents (CAST) appear in regions where FLLW2 is low. Here, a width of a signal reflects length of time during which an optical signal is being detected on a pulse-like signal waveform where the vertical axis represents signal intensity and the horizontal axis represents time.

DETD With another result of measurements of bacteria, scattergrams are generated from forward-scattered light signal and fluorescence signal. FIG. 11(e) is a scattergram in which the horizontal axis represents fluorescence intensity (B-FLH) and the vertical axis represents forward-scattered light intensity (high-sensitivity) (B-FSC). In urinary particle measurements, as shown by the scattergram in FIG. 11(c), a range of bacteria appearance overlaps with that of mucus fibril (MUCUS), yeast-like fungi (YLC), and sperms (SPERM). However, with bacteria measurement, foreign substances such as mucus fibril and debris of erythrocytes are caused to constrict by a bacteria measurement reagent, and therefore, there is such a region where only bacteria appear independently. In addition, since measurements are made with approximately 10 times improved sensitivity compared to urinary particle measurements, small-sized bacteria can also be detected with high accuracy, thereby ensuring accurate results of bacteria measurements.

DETD FIG. 11(e) shows a standard appearance region of bacteria (BACT), while the appearance region is depending on types of bacteria. FIG. 13 is an example of measurements of a sample in which a large quantity of cocci appeared and chained. In this scattergram, the region where bacteria (BACT) appeared is distributed with an angle of approximately 45° with regard to the horizontal axis (fluorescence intensity). In other words, bacteria (BACT) appeared in regions where forward-scattered light intensity (FSC) is high. With such samples, in the urinary particle measurement (SED), bacteria would appear in wider ranges, and eventually appear even in ranges, of erythrocyte appearance regions, where forward-scattered light intensity (FSC) is low. With these samples, reliability of erythrocyte measurement is low. Meanwhile, FIG. 12 shows an example of measurement of samples containing bacilli. In this scattergram, the region where bacteria (BACT) appeared is distributed with a lower angle (approximately 5 to 10 degrees) with regard to the horizontal axis (fluorescence intensity). Namely, bacteria (BACT) appeared in a region where forward-scattered light intensity (FSC) is low. With such specimens, even if bacilli is contained in a large amount,

forward-scattered light intensity (FSC) in the bacteria appearance region is lower than that of the erythrocyte appearance region, and erythrocyte measurement is not affected by bacteria. Similarly, influences of bacteria on leukocyte (WBC) appearance region in the urinary particle (SED) measurement can be confirmed from bacteria distribution in the bacteria measurement (BAC). Judgment of presence or absence of influences on measurement results of other particles according to the tendency of bacteria distribution as mentioned above is carried out by algorithm analysis by the personal computer 13 (analysis section), and results of judgment are displayed on the display together with other measurement results in the Step S39.

DETD In the present embodiment, a scattergram is generated in measurement of particles. However, the scattergram need not necessarily be generated. The scattergram generated by the apparatus for analyzing particles in urine U is a distribution map in which a plurality of parameters extracted from signal data corresponding to each particles in urine are used as coordinate axes. The scattergram is generated as one technique of algorithm analysis. One advantage of the scattergram is that a user can visually confirm results of measurements. However, so far as signal data corresponding to each particle are used for analysis, classification and counting of particles are possible without necessarily generating a scattergram. The only thing that has to be determined is in what range signal data corresponding to each particle should be maintained and then into what type of particle the particle should be classified. In this specification, regardless of necessity of generation of a scattergram, a range of distribution of data obtained from each particle is referred to as an appearance region.

L9 ANSWER 10 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2007:62904 USPATFULL <<LOGINID::20090129>>  
 TITLE: HLA binding motifs and peptides and their uses  
 INVENTOR(S): Grey, Howard M., La Jolla, CA, UNITED STATES  
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 Kast, W. Martin, La Canada, CA, UNITED STATES

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PATENT INFORMATION:	US 20070055049	A1	20070308
APPLICATION INFO.:	US 2004-817970	A1	20040406 (10)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 1997-821739, filed on 20 Mar 1997, ABANDONED Continuation-in-part of Ser. No. US 1996-589107, filed on 23 Jan 1996, ABANDONED Continuation-in-part of Ser. No. US 1995-451913, filed on 26 May 1995, ABANDONED Continuation-in-part of Ser. No. US 1994-186266, filed on 25 Jan 1994, GRANTED, Pat. No. US 5662907 Continuation-in-part of Ser. No. US 1993-159339, filed on 29 Nov 1993, GRANTED, Pat. No. US 6037135 Continuation-in-part of Ser. No. US 1993-103396, filed on 6 Aug 1993, ABANDONED Continuation-in-part of Ser. No. US 1993-27746, filed on 5 Mar 1993, ABANDONED Continuation-in-part of Ser. No. US 1992-926666, filed on 7 Aug 1992, ABANDONED Continuation-in-part of Ser. No. US 1994-347610, filed on 1 Dec 1994, ABANDONED Continuation-in-part of Ser. No. US 1993-159339, filed on 29 Nov 1993, GRANTED, Pat.		

No. US 6037135 Continuation-in-part of Ser. No. US 1993-103396, filed on 6 Aug 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1993-27746, filed on 5 Mar 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1992-926666, filed on 7 Aug 1992, ABANDONED  
Continuation-in-part of Ser. No. US 2000-665510, filed on 19 Sep 2000, ABANDONED  
Continuation-in-part of Ser. No. US 1994-347610, filed on 1 Dec 1994, ABANDONED  
Continuation-in-part of Ser. No. US 1993-159339, filed on 29 Nov 1993, GRANTED, Pat. No. US 6037135  
Continuation-in-part of Ser. No. US 1993-103396, filed on 6 Aug 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1993-27746, filed on 5 Mar 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1992-926666, filed on 7 Aug 1992, ABANDONED  
Continuation-in-part of Ser. No. US 1998-17524, filed on 3 Feb 1998, ABANDONED  
Continuation-in-part of Ser. No. US 1996-589107, filed on 23 Jan 1996, ABANDONED  
Continuation-in-part of Ser. No. US 1996-758409, filed on 27 Nov 1996, ABANDONED  
Continuation-in-part of Ser. No. US 1997-821739, filed on 20 Mar 1997, ABANDONED  
Continuation-in-part of Ser. No. US 1996-589107, filed on 23 Jan 1996, ABANDONED  
Continuation-in-part of Ser. No. US 1995-451913, filed on 26 May 1995, ABANDONED  
Continuation-in-part of Ser. No. US 1994-347610, filed on 1 Dec 1994, ABANDONED  
Continuation-in-part of Ser. No. US 1993-159339, filed on 29 Nov 1993, GRANTED, Pat. No. US 6037135  
Continuation-in-part of Ser. No. US 1993-103396, filed on 6 Aug 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1993-27746, filed on 5 Mar 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1992-926666, filed on 7 Aug 1992, ABANDONED  
Continuation-in-part of Ser. No. US 1994-186266, filed on 25 Jan 1994, GRANTED, Pat. No. US 5662907  
Continuation-in-part of Ser. No. US 1993-159339, filed on 29 Nov 1993, GRANTED, Pat. No. US 6037135  
Continuation-in-part of Ser. No. US 1993-103396, filed on 6 Aug 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1993-27746, filed on 5 Mar 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1992-926666, filed on 7 Aug 1992, ABANDONED  
Continuation-in-part of Ser. No. US 1998-17735, filed on 3 Feb 1998, ABANDONED  
Continuation-in-part of Ser. No. US 1994-205713, filed on 4 Mar 1994, ABANDONED  
Continuation-in-part of Ser. No. US 1993-159184, filed on 29 Nov 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1993-73205, filed on 4 Jun 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1993-27146, filed on 5 Mar 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1996-589108, filed on 23 Jan 1996, ABANDONED  
Continuation-in-part of Ser. No. US 1995-454033, filed on 26 May 1995, ABANDONED  
Continuation-in-part of Ser. No. US 1994-349177, filed on 2 Dec 1994, ABANDONED  
Continuation-in-part of Ser. No. US 1993-159184, filed on 29 Nov 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1993-73205, filed on 4 Jun 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1993-27146, filed on 5 Mar 1993, ABANDONED  
Continuation-in-part of Ser. No. US 1997-822382, filed on 20 Mar 1997, ABANDONED  
Continuation-in-part of Ser. No. US 1996-753622, filed on 27 Nov 1996, ABANDONED  
Continuation-in-part of Ser. No. US 1994-205713, filed on 4 Mar 1994, ABANDONED

Continuation-in-part of Ser. No. US 1993-159184, filed on 29 Nov 1993, ABANDONED Continuation-in-part of Ser. No. US 1993-73205, filed on 4 Jun 1993, ABANDONED Continuation-in-part of Ser. No. US 1993-27146, filed on 5 Mar 1993, ABANDONED Continuation-in-part of Ser. No. US 1995-454033, filed on 26 May 1995, ABANDONED Continuation-in-part of Ser. No. US 1994-349177, filed on 2 Dec 1994, ABANDONED Continuation-in-part of Ser. No. US 1993-159184, filed on 29 Nov 1993, ABANDONED Continuation-in-part of Ser. No. US 1993-73205, filed on 4 Jun 1993, ABANDONED Continuation-in-part of Ser. No. US 1993-27146, filed on 5 Mar 1993, ABANDONED Continuation-in-part of Ser. No. US 1998-17743, filed on 3 Feb 1998, ABANDONED Continuation-in-part of Ser. No. US 1996-753615, filed on 27 Nov 1996, ABANDONED Continuation-in-part of Ser. No. US 1996-590298, filed on 23 Jan 1996, ABANDONED Continuation-in-part of Ser. No. US 1995-452843, filed on 30 May 1995, ABANDONED Continuation-in-part of Ser. No. US 1994-344824, filed on 23 Nov 1994, ABANDONED Continuation-in-part of Ser. No. US 1994-278634, filed on 21 Jul 1994, ABANDONED Continuation-in-part of Ser. No. US 1995-452843, filed on 30 May 1995, ABANDONED Continuation-in-part of Ser. No. US 1994-344824, filed on 23 Nov 1994, ABANDONED Continuation-in-part of Ser. No. US 1994-278634, filed on 21 Jul 1994, ABANDONED Continuation-in-part of Ser. No. US 1994-344824, filed on 23 Nov 1994, ABANDONED Continuation-in-part of Ser. No. US 1994-278634, filed on 21 Jul 1994, ABANDONED Continuation-in-part of Ser. No. US 1999-226775, filed on 6 Jan 1999, ABANDONED Continuation-in-part of Ser. No. US 1997-815396, filed on 10 Mar 1997, ABANDONED Continuation-in-part of Ser. No. US 1995-485218, filed on 7 Jun 1995, ABANDONED Continuation-in-part of Ser. No. US 1994-305871, filed on 14 Sep 1994, GRANTED, Pat. No. US 5736142 Continuation-in-part of Ser. No. US 1993-121101, filed on 14 Sep 1993, ABANDONED Continuation-in-part of Ser. No. US 2002-30014, filed on 24 Jul 2002, ABANDONED A 371 of International Ser. No. WO 2000-US17842, filed on 28 Jun 2000 Continuation-in-part of Ser. No. US 2002-121415, filed on 11 Apr 2002, PENDING Continuation-in-part of Ser. No. US 1998-189702, filed on 10 Nov 1998, PENDING Continuation-in-part of Ser. No. US 1998-98584, filed on 17 Jun 1998, ABANDONED Continuation-in-part of Ser. No. WO 2003-US31308, filed on 3 Oct 2003, PENDING Continuation-in-part of Ser. No. US 1999-260714, filed on 1 Mar 1999, ABANDONED Continuation-in-part of Ser. No. US 2004-470364, filed on 9 Apr 2004, PENDING A 371 of International Ser. No. WO 2002-US2708, filed on 29 Jan 2002 Continuation-in-part of Ser. No. US 2001-935476, filed on 22 Aug 2001, ABANDONED Continuation-in-part of Ser. No. US 1999-346105, filed on 30 Jun 1999, ABANDONED Continuation-in-part of Ser. No. US 1999-469201, ABANDONED A 371 of International Ser. No. WO 2001-US51650, filed on 18 Oct 2001

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PRIORITY INFORMATION:	US 1996-13833P	19960321 (60)
	US 1996-13833P	19960321 (60)

US 1996-13980P 19960321 (60)  
US 1996-13113P 19960311 (60)  
US 1999-141422P 19990629 (60)  
US 2002-416207P 20021003 (60)  
US 2002-417269P 20021008 (60)  
US 2001-264969P 20010129 (60)  
US 2001-285624P 20010420 (60)  
US 2000-242350P 20001019 (60)

DOCUMENT TYPE: Utility  
FILE SEGMENT: APPLICATION  
LEGAL REPRESENTATIVE: STERNE, KESSLER, GOLDSTEIN & FOX PLLC, 1100 NEW YORK  
AVENUE, N.W., WASHINGTON, DC, 20005, US

NUMBER OF CLAIMS: 2  
EXEMPLARY CLAIM: 1  
NUMBER OF DRAWINGS: 57 Drawing Page(s)  
LINE COUNT: 11380

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DRWD FIG. 45 shows a scattergram of the log of relative binding  
plotted against the "Grouped Ratio" algorithm for 9 mer peptides.

DRWD FIG. 46 shows a scattergram of the log of relative binding  
plotted against the average "Log of Binding" algorithm score for 9 mer  
peptides.

DRWD FIG. 47 and FIG. 48 show scattergrams of a set of 10-mer  
peptides containing preferred residues in positions 2 and 10 as scored  
by the "Grouped Ratio" and "Log of Binding" algorithms.

DETD For therapeutic or prophylactic immunization purposes, the peptides of  
the invention can be expression vectors include attenuated viral hosts,  
such as vaccinia or fowlpox. This approach involves the use of vaccinia  
virus, for example, as a vector to express nucleotide sequences that  
encode the peptides of the invention. Upon introduction into an acutely  
or chronically infected host or into a non-infected host, the  
recombinant vaccinia virus expresses the immunogenic peptide, and  
thereby elicits a host CTL and/or HTL response. Vaccinia vectors and  
methods useful in immunization protocols are described in, e.g., U.S.  
Pat. No. 4,722,848. Another vector is BCG (Bacille Calmette  
Guerin). BCG vectors are described in Stover et al., Nature 351:456-460  
(1991). A wide variety of other vectors useful for therapeutic  
administration or immunization of the peptides of the invention, e.g.  
adeno and adeno-associated virus vectors, retroviral vectors, Salmonella  
typhi vectors, detoxified anthrax toxin vectors, and the like, will be  
apparent to those skilled in the art from the description herein.

DETD For therapeutic or immunization purposes, the peptides of the invention  
can also be expressed by vectors. Examples of expression vectors include  
attenuated viral hosts, such as vaccinia or fowlpox. This approach  
involves the use of vaccinia virus as a vector to express nucleotide  
sequences that encode the peptides of the invention. Upon introduction  
into an acutely or chronically infected host or into a non-infected  
host, the recombinant vaccinia virus expresses the immunogenic peptide,  
and thereby elicits a host CTL response. Vaccinia vectors and methods  
useful in immunization protocols are described in, e.g., U.S. Pat. No.  
4,722,848. Another vector is BCG (Bacille Calmette Guerin).  
BCG vectors are described in Stover, et al. Nature 351:456-60 (1991). A  
wide variety of other vectors useful for therapeutic administration or  
immunization of the peptides of the invention, e.g., Salmonella typhi  
vectors, retroviral vectors, adenoviral or adeno-associated viral  
vectors, and the like will be apparent to those skilled in the art from  
the description herein.

DETD For therapeutic or immunization purposes, the peptides of the invention  
can also be expressed by attenuated viral hosts, such as vaccinia or  
fowlpox. This approach involves the use of vaccinia virus as a vector to  
express nucleotide sequences that encode the peptides of the invention.

Upon introduction into an acutely or chronically infected host or into a noninfected host, the recombinant vaccinia virus expresses the immunogenic peptide, and thereby elicits a host CTL response. Vaccinia vectors and methods useful in immunization protocols are described in, e.g., U.S. Pat. No. 4,722,848, incorporated herein by reference. Another vector is BCG (Bacille Calmette Guerin). BCG vectors are described in Stover et al. (Nature 351:456-60 (1991)) which is incorporated herein by reference. A wide variety of other vectors useful for therapeutic administration or immunization of the peptides of the invention, e.g., Salmonella typhi vectors and the like, will be apparent to those skilled in the art from the description herein.

DETD In the present "Grouped Ratio" algorithm residues have been grouped by similarity. This avoids the problem encountered with some rare residues, such as tryptophan, where there are too few occurrences to obtain a statistically significant ratio. TABLES 165 and 166 is a listing of scores obtained by grouping for each of the twenty amino acids by position for 9-mer peptides containing perfect 2/9 motifs. A peptide is scored in the "Grouped Ratio" algorithm as a product of the scores of each of its residues. In the case of positions other than 2 and 9, the scores have been derived using a set of peptides which contain only preferred residues in positions 2 and 9. To enable us to extend our "Grouped Ratio" algorithm to peptides which may have residues other than the preferred ones at 2 and 9, scores for 2 and 9 have been derived from a set of peptides which are single amino acid substitutions at positions 2 and 9. FIG. 45 shows a scattergram of the log of relative binding plotted against "Grouped Ratio" algorithm score for our collection of 9-mer peptides from the previous example.

DETD An algorithm using the average binding affinity of all the peptides with a certain amino acid (or amino acid type) at a certain position has the advantage of including all of the peptides in the analysis, and not just good/intermediate binders and non-binders. Moreover, it gives a more quantitative measure of affinity than the simpler "Grouped Ratio" algorithm. We have created such an algorithm by calculating for each amino acid, by position, the average log of binding when that particular residue occurs in our set of 160 2.9 motif containing peptides. These values are shown in TABLE 168. The algorithm score for a peptide is then taken as the sum of the scores by position for each residues. FIG. 46 shows a scattergram of the log of relative binding against the average "Log of Binding" algorithm score. TABLE 167 shows the ability of the two algorithms to predict peptide binding at various levels, as a function of the cut-off score used. The ability of a 2.9 motif to predict binding in the same peptide set is also shown for reference purposes. It is clear from this comparison that both algorithms of this invention have a greater ability to predict populations with higher frequencies of good binders than a 2.9 motif alone. Differences between the "Grouped Ratio" algorithm and the "Log of Binding" algorithm are small in the set of peptides analyzed here, but do suggest that the "Log of Binding" algorithm is a better, if only slightly, predictor than the "Grouped Ratio" algorithm.

DETD Using the methods described in the proceeding example, an analogous set of algorithms has been developed for predicting the binding of 10-mer peptides. TABLE 169 shows the scores used in a "Grouped Ratio" algorithm, and TABLE 170 shows the "Log of Binding" algorithm scores, for 10-mer peptides. TABLE 171 shows a comparison of the application of the two different algorithmic methods for selecting binding peptides. FIG. 47 and FIG. 48 show, respectively, scattergrams of a set of 10-mer peptides containing preferred residues in positions 2 and 10 as scored by the "Grouped Ratio" and "Log of Binding" algorithms.



TITLE: ILT3 binding molecules and uses therefor  
 INVENTOR(S): Ponath, Paul, San Francisco, CA, UNITED STATES  
 Rao, Patricia, Acton, MA, UNITED STATES  
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 Ponte, Jose F., South Boston, MA, UNITED STATES  
 PATENT ASSIGNEE(S): TolerRx, Inc., Cambridge, MA, UNITED STATES (U.S. corporation)

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PATENT INFORMATION:	US 20070041982	A1	20070222
APPLICATION INFO.:	US 2006-471397	A1	20060619 (11)

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PRIORITY INFORMATION:	US 2005-691912P	20050617 (60)
	US 2005-723340P	20051004 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	LAHIVE & COCKFIELD, LLP, ONE POST OFFICE SQUARE, BOSTON, MA, 02109-2127, US	
NUMBER OF CLAIMS:	43	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	6 Drawing Page(s)	
LINE COUNT:	3257	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD E. coli is one prokaryotic host particularly useful for cloning the polynucleotides (e.g., DNA sequences) of the present invention. Other microbial hosts suitable for use include bacilli, such as *Bacillus subtilis*, and other enterobacteriaceae, such as *Salmonella*, *Serratia*, and various *Pseudomonas* species. In these prokaryotic hosts, one can also make expression vectors, which will typically contain expression control sequences compatible with the host cell (e.g., an origin of replication). In addition, any number of a variety of well-known promoters will be present, such as the lactose promoter system, a tryptophan (*trp*) promoter system, a beta-lactamase promoter system, or a promoter system from phage lambda. The promoters will typically control expression, optionally with an operator sequence, and have ribosome binding site sequences and the like, for initiating and completing transcription and translation.

DETD Typical antigens of interest may be classified as follows: protein antigens, such as ceruloplasmin and serum albumin; bacterial antigens, such as teichoic acids, flagellar antigens, capsular polysaccharides, and extra-cellular bacterial products and toxins; glycoproteins and glycolipids; viruses, such as animal, plant, and bacterial viruses; conjugated and synthetic antigens, such as proteinhaptens conjugates, molecules expressed preferentially by tumors, compared to normal tissue; synthetic polypeptides; and nucleic acids, such as ribonucleic acid and deoxyribonucleic acid. The term "infectious agent," as used herein, includes any agent which expresses an antigen which elicits a host cellular immune response. Non-limiting examples of viral antigens which may be considered useful as include, but are not limited to, the nucleoprotein (NP) of influenza virus and the Gag proteins of HIV. Other heterologous antigens include, but are not limited to, HIV Env protein or its component parts gp120 and gp41, HIV Nef protein, and the HIV Pol proteins, reverse transcriptase and protease. In addition, other viral antigens such as Ebola virus (EBOV) antigens, such as, for example, EBOV NP or glycoprotein (GP), either full-length or GP deleted in the mucin region of the molecule (Yang Z-Y, et al. (2000) Nat Med 6:886-9, 2000), small pox antigens, hepatitis A, B or C virus, human rhinovirus such as

type 2 or type 14, Herpes simplex virus, poliovirus type 2 or 3, foot-and-mouth disease virus (FMDV), rabies virus, rotavirus, influenza virus, coxsackie virus, human papilloma virus (HPV), for example the type 16 papilloma virus, the E7 protein thereof, and fragments containing the E7 protein or its epitopes; and simian immunodeficiency virus (SIV) may be used. The antigens of interest need not be limited to antigens of viral origin. Parasitic antigens, such as, for example, malarial antigens are included, as are fungal antigens, bacterial antigens and tumor antigens. Examples of antigens derived from bacteria are those derived from *Bordetella pertussis* (e.g., P69 protein and filamentous haemagglutinin (FHA) antigens), *Vibrio cholerae*, *Bacillus anthracis*, and *E. coli* antigens such as *E. coli* heat Labile toxin B subunit (LT-B), *E. coli* K88 antigens, and enterotoxigenic *E. coli* antigens. Other examples of antigens include *Schistosoma mansoni* P28 glutathione S-transferase antigens (P28 antigens) and antigens of flukes, mycoplasma, roundworms, tapeworms, *Chlamydia trachomatis*, and malaria parasites, e.g., parasites of the genus *plasmodium* or babesia, for example *Plasmodium falciparum*, and peptides encoding immunogenic epitopes from the aforementioned antigens.

DETD An infection, disease or disorder which may be treated or prevented by the administration of a composition of the invention includes any infection, disease or disorder wherein a host immune response acts to prevent the infection, disease or disorder. Diseases, disorders, or infection which may be treated or prevented by the administration of the compositions of the invention include, but are not limited to, any infection, disease or disorder caused by or related to a fungus, parasite, virus, or bacteria, diseases, disorders or infections caused by or related to various agents used in bioterrorism, listeriosis, Ebola virus, SARS, small pox, hepatitis A, hepatitis B, hepatitis C, diseases and disorders caused by human rhinovirus, HIV and AIDS, Herpes, polio, foot-and-mouth disease, rabies, diseases or disorders caused by or related to: rotavirus, influenza, coxsackie virus, human papilloma virus, SIV, malaria, cancer, e.g., tumors, and diseases or disorders caused by or related to infection by *Bordetella pertussis*, *Vibrio cholerae*, *Bacillus anthracis*, *E. coli*, flukes, mycoplasma, roundworms, tapeworms, *Chlamydia trachomatis*, and malaria parasites, etc.

DETD As used herein, the term "bacterial infections" include infections with a variety of bacterial organisms, including gram-positive and gram-negative bacteria. Examples include, but are not limited to, *Neisseria* spp, including *N. gonorrhea* and *N. meningitidis*, *Streptococcus* spp, including *S. pneumoniae*, *S. pyogenes*, *S. agalactiae*, *S. mutans*; *Haemophilus* spp, including *H. influenzae* type B, non typeable *H. influenzae*, *H. ducreyi*; *Moraxella* spp, including *M. catarrhalis*, also known as *Branhamella catarrhalis*; *Bordetella* spp, including *B. pertussis*, *B. parapertussis* and *B. bronchiseptica*; *Mycobacterium* spp., including *M. tuberculosis*, *M. bovis*, *M. leprae*, *M. avium*, *M. paratuberculosis*, *M. smegmatis*; *Legionella* spp, including *L. pneumophila*; *Escherichia* spp, including enterotoxigenic *E. coli*, enterohemorrhagic *E. coli*, enteropathogenic *E. coli*; *Vibrio* spp, including *V. cholera*, *Shigella* spp, including *S. sonnei*, *S. dysenteriae*, *S. flexnerii*; *Yersinia* spp, including *Y. enterocolitica*, *Y. pestis*, *Y. pseudotuberculosis*, *Campylobacter* spp, including *C. jejuni* and *C. coli*; *Salmonella* spp, including *S. typhi*, *S. paratyphi*, *S. choleraesuis*, *S. enteritidis*; *Listeria* spp., including *L. monocytogenes*; *Helicobacter* spp, including *H. pylori*; *Pseudomonas* spp, including *P. aeruginosa*, *Staphylococcus* spp., including *S. aureus*, *S. epidermidis*; *Enterococcus* spp., including *E. faecalis*, *E. faecium*; *Clostridium* spp., including *C. tetani*, *C. botulinum*, *C. difficile*; *Bacillus* spp., including *B. anthracis*; *Corynebacterium* spp., including *C. diphtheriae*; *Borrelia* spp., including *B. burgdorferi*, *B. garinii*, *B. afzelii*, *B. andersonii*,

B. hermsii; Ehrlichia spp., including E. equi and the agent of the Human Granulocytic Ehrlichiosis; Rickettsia spp., including R. rickettsii; Chlamydia spp., including C. trachomatis, C. pneumoniae, C. psittaci; Leptospira spp., including L. interrogans; Treponema spp., including T. pallidum, T. denticola, T. hyodysenteriae. Preferred bacteria include, but are not limited to, Listeria, mycobacteria, mycobacteria (e.g., tuberculosis), Anthrax, Salmonella and Listeria monocytogenes.

DETD Intracellular calcium flux studies using flow cytometry analysis was performed as described by Rabin, et al. (J Immunol. (1999)162:3840-3850). Briefly, monocyte-derived dendritic cells (2+10.sup.7) were suspended in HBSS-HEPES (HBSS supplemented with 10 mM HEPES, Ca.sup.++, Mg.sup.++, and 1% fetal calf serum). Indo-1 and pleuronic detergent (Molecular Probes, Eugene, Oreg.) were added at final concentrations of 5 µM and 300 µg/mL, respectively. The cell suspension was incubated at 30° C. for 45 minutes with gentle agitation. Cells were then washed twice with the HBSS-HEPES, stained with anti-CD1a, and washed again. Calcium flux for CD1a.sup.+ dendritic cells was performed using a FACSVantage flow cytometer (Becton Dickinson) equipped with an argon laser tuned to 488 nM and a krypton laser tuned to 360 nM. Indo-1 fluorescence was analyzed at 390/20 nM and 530/20 nM for bound and free calcium, respectively. Before stimulation, cell suspensions were warmed at 37° C. for 3 minutes. The CD1a.sup.+ cell population was gated, and baseline fluorescent ratios were collected for 30 seconds. Cells were then stimulated with either fMLP (10.sup.-5 M), T-20 peptide (10.sup.-5 M), or F-peptide (10.sup.-5 M) followed by fMLP (10.sup.-8 M). Collections continued until calcium flux returned to basal levels. Changes in Indo-1 fluorescence were expressed as the ratio of bound to free intracellular calcium, and scattergrams represented the entire CD1a.sup.+ cell population at the time of stimulation. Data analysis was performed using Flowjo software (Tree Star, San Carlos, Calif.).

L9 ANSWER 12 OF 42 USPATFULL on STN

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 TITLE: NOEY2 gene compositions and methods of use  
 INVENTOR(S): Yu, Yinhua, Pearland, TX, UNITED STATES  
 Xu, Fengji, Houston, TX, UNITED STATES  
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 PATENT ASSIGNEE(S): Board of Regents, the University of Texas System,  
 Austin, TX, UNITED STATES (U.S. corporation)

	NUMBER	KIND	DATE
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CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Patient outcome may be characterized by one of the time-to-event variables 1) survival time or 2) disease-free survival (DFS) or by the

binary variable indicating either 3) response or 4) partial response to chemotherapy (Cox, 1972; Modern Applied Statistics with S-Plus, 1994; Grambsch and Therneau, 1994; Harrington and Fleming, 1991). Each of these evaluations may be carried out by regression analysis, with the patient outcome as the response variable in the regression model and NOEY2 included as a predictive covariate along with the established predictors disease stage, disease grade, amount of residual disease post surgery, and other molecular markers, including HER-2, EGFR, fms, and p53. Because NOEY2 is recorded as an ordinal variable taking on the values 0,1,2,3 or 4, it may be evaluated first as a numerical covariate and subsequently as a categorical covariate in each regression analysis. For each patient outcome, specific questions to be addressed include whether NOEY2 per se is predictive, if so whether the effect of NOEY2 on patient outcomes survival of DFS changes over time, and whether any significant effect of NOEY2 persists in the presence of the established predictors noted above. Relationships between pairs of covariates may be evaluated by computing standard Pearson correlations and Spearman rank correlations between numerical variables and constructing their smoothed scattergrams, by cross-tabulating categorical variables, and by carrying out Kruskal-Wallis or Wilcoxon-Mann-Whitney tests to assess the change of a numerical variable across a categorical variable.

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L9 ANSWER 13 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2006:340908 USPATFULL <<LOGINID::20090129>>

TITLE: CaR receptor as a mediator of migratory cell chemotaxis and/or chemokinesis

INVENTOR(S): Poznansky, Mark C., Charlestown, MA, UNITED STATES  
Brown, Edward M., Milton, MA, UNITED STATES  
Scadden, David T., Weston, MA, UNITED STATES  
Olszak, Ivona T., Charlestown, MA, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20060292689	A1	20061228
APPLICATION INFO.:	US 2006-429902	A1	20060508 (11)
RELATED APPLN. INFO.:	Division of Ser. No. US 2001-2854, filed on 1 Nov 2001, PENDING Continuation-in-part of Ser. No. WO 2000-US15440, filed on 2 Jun 2000, PENDING		

	NUMBER	DATE
PRIORITY INFORMATION:	US 2000-200861P	20000501 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	EDWARDS & ANGELL, LLP, P.O. BOX 55874, BOSTON, MA, 02205, US	
NUMBER OF CLAIMS:	64	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	6 Drawing Page(s)	
LINE COUNT:	2783	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DRWD FIG. 1(a): Scattergram showing CaR positive stain on CD14.sup.+ monocytes (upper panel), and inhibition of anti-CaR antibody binding to CaR by preincubating CD14.sup.+ monocytes with CaR peptide (lower panel); FIG. 1(b) Graphs showing elevation of CD14.sup.+ intracellular Ca.sup.++ concentration following elevation in the extracellular Ca.sup.++ concentration or addition of the selective CaR activator NPS R-467 in the extracellular medium.

DETD Bacteria in general include but are not limited to: *P. aeruginosa*; *Bacillus anthracis*; *E. coli*, *Enterocytozoon bieneusi*; *Klebsiella* sp.; *Klebsiella pneumoniae*; *Serratia* sp.; *Pseudomonas* sp.; *P. cepacia*; *Acinetobacter* sp.; *S. epidermis*; *E. faecalis*; *S. pneumoniae*; *S. aureus*; *Haemophilus* sp.; *Haemophilus Influenza*; *Neisseria* Sp.; *Neisseria gonorrhoeae*; *Neisseria meningitis*; *Helicobacter pylori*; *Bacteroides* sp.; *Citrobacter* sp.; *Branhamella* sp.; *Salmonella* sp.; *Salmonella typhi*;

Shigella sp.; S. pyogenes; Proteus sp.; Clostridium sp.; Erysipelothrix sp.; Listeria sp.; Pasteurella multocida; Streptobacillus sp.; Spirillum sp.; Fusospirocheta sp.; Actinomycetes; Mycoplasma sp.; Chlamydiae sp.; Chlamydia trachomatis; Campylobacter jejuni; Cyclospora cayatanensis; Rickettsia sp.; Spirochaeta, including Treponema pallidum and Borrelia sp.; Legionella sp.; Legionella pneumophila; Mycobacteria sp.; Mycobacterium tuberculosis; Ureaplasma sp.; Streptomyces sp.; Trichomonas sp.; and P. mirabilis, as well as toxins, that include, but are not limited to, Anthrax toxin (EF); Adenylate cyclase toxin; Cholera enterotoxin; E. coli LT toxin; Escherichia coli 0157:H7; Shiga toxin; Botulinum Neurotoxin Type A heavy and light chains; Botulinum Neurotoxin Type B heavy and light chains; Tetanus toxin; Tetanus toxin C fragment; Diphtheria toxin; Pertussis toxin; Parvovirus B19; Staphylococcus enterotoxins; Toxic shock syndrome toxin (TSST-1); Erythrogenic toxin; and Vibrio cholerae 0139.

DETD FIG. 1a: CD14<sup>sup</sup> monocytes stain positively for the CaR, and binding of anti-CaR antibody is inhibitable by preincubation with CaR peptide. Purified peripheral blood CD14<sup>+</sup> monocytes (scattergram) were exposed to anti-CaR antibody (solid area in histogram) or isotype control (open area) and examined by flow cytometry. Monocytes were also preincubated with CaR peptide prior to staining with anti-CaR antibody (dashed area). Data represent one of ten independent experiments with comparable results.

L9 ANSWER 14 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2006:288016 USPATFULL <<LOGINID::20090129>>

TITLE: Antibody variants and uses thereof

INVENTOR(S): Adams, Camellia W., San Jose, CA, UNITED STATES

Lowman, Henry B., El Granada, CA, UNITED STATES

Nakamura, Gerald R., San Francisco, CA, UNITED STATES

PATENT ASSIGNEE(S): Genentech, Inc., South San Francisco, CA, UNITED STATES (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20060246004	A1	20061102
APPLICATION INFO.:	US 2006-348609	A1	20060206 (11)

	NUMBER	DATE
PRIORITY INFORMATION:	US 2005-702571P	20050725 (60)
	US 2005-689404P	20050610 (60)
	US 2005-651111P	20050207 (60)

DOCUMENT TYPE: Utility

FILE SEGMENT: APPLICATION

LEGAL REPRESENTATIVE: GENENTECH, INC., 1 DNA WAY, SOUTH SAN FRANCISCO, CA, 94080, US

NUMBER OF CLAIMS: 58

EXEMPLARY CLAIM: 1

NUMBER OF DRAWINGS: 14 Drawing Page(s)

LINE COUNT: 6158

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Expression and cloning vectors may contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g., the gene encoding D-alanine racemase for Bacilli.

DETD Suitable host cells for cloning or expressing the DNA in the vectors herein are the prokaryote, yeast, or higher eukaryote cells described

above. Suitable prokaryotes for this purpose include eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as Escherichia, e.g., E. coli, Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. licheniformis (e.g., B. licheniformis 41P disclosed in DD 266,710 published 12 Apr. 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. One preferred E. coli cloning host is E. coli 294 (ATCC 31,446), although other strains such as E. coli B, E. coli X1776 (ATCC 31,537), and E. coli W3110 (ATCC 27,325) are suitable. These examples are illustrative rather than limiting.

DETD Peripheral B-cell concentrations were determined by a FACS method that counted CD3-/CD40+ cells. The percent of CD3-CD40+B cells of total lymphocytes in monkey samples were obtained by the following gating strategy. The lymphocyte population was marked on the forward scatter/side scatter scattergram to define Region 1 (R1). Using events in R1, fluorescence intensity dot plots were displayed for CD40 and CD3 markers. Fluorescently labeled isotype controls were used to determine respective cutoff points for CD40 and CD3 positivity.

L9 ANSWER 15 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2006:233777 USPATFULL <<LOGINID::20090129>>

TITLE: Detection of activation of endothelial cells as surrogate marker for angiogenesis

INVENTOR(S): Moore, Sean C., Durham, NC, UNITED STATES  
Singh, Sharat, Los Altos, CA, UNITED STATES  
Salimi-Moosavi, Hossein, Sunnyvale, CA, UNITED STATES  
Cao, Liching, Vallejo, CA, UNITED STATES  
Sperinde, Jeff, El Granada, CA, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20060199231	A1	20060907
APPLICATION INFO.:	US 2005-267870	A1	20051103 (11)

	NUMBER	DATE
PRIORITY INFORMATION:	US 2004-625694P	20041104 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	JONES DAY, 222 EAST 41ST ST, NEW YORK, NY, 10017, US	
NUMBER OF CLAIMS:	37	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	26 Drawing Page(s)	
LINE COUNT:	4182	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM Laser scanning cytometry (LSC) has also been used for quantitative analysis of antiangiogenic activity in clinical studies. In LSC measurement automated lasers detect individual cells within the mapped region of tumor biopsy samples based on multicolor immunofluorescence staining of biomarkers. Each cell is plotted on a scattergram based on its relative fluorescence intensity. LSC-generated scattergrams display the percentage of cell populations, for example, apoptotic endothelial cells. Alternative, cellular protein expression levels, e.g., phosphorylated VEGF receptor-2, may be measured by histogram analysis. See review by Davis et al. (2003) Br. J. Cancer 89:8-14.

DETD Other immuno-modulating agents other than cytokines may also be used in conjunction with CPT and a COX-2 inhibitor to inhibit abnormal cell

growth. Examples of such immuno-modulating agents include, but are not limited to bacillus Calmette-Guerin, levamisole, and octreotide, a long-acting octapeptide that mimics the effects of the naturally occurring hormone somatostatin.

DETD An adjuvant may be used to augment the immune response to TAAs. Examples of adjuvants include, but are not limited to, bacillus Calmette-Guerin (BCG), endotoxin lipopolysaccharides, keyhole limpet hemocyanin (GKLH), interleukin-2 (IL-2), granulocyte-macrophage colony-stimulating factor (GM-CSF) and cytoxan, a chemotherapeutic agent which is believe to reduce tumor-induced suppression when given in low doses.

L9 ANSWER 16 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2006:158579 USPATFULL <<LOGINID::20090129>>  
 TITLE: G protein coupled receptors and uses thereof  
 INVENTOR(S): Gaitanaris, George A., Seattle, WA, UNITED STATES  
 Bergmann, John E., Mercer Island, WA, UNITED STATES  
 Gragerov, Alexander, Seattle, WA, UNITED STATES  
 Hohmann, John, La Conner, WA, UNITED STATES  
 Li, Fusheng, Seattle, WA, UNITED STATES  
 Madisen, Linda, Seattle, WA, UNITED STATES  
 McIlwain, Kellie L., Renton, WA, UNITED STATES  
 Pavlova, Maria N., Seattle, WA, UNITED STATES  
 Vassilatis, Demitri, Seattle, WA, UNITED STATES  
 Zeng, Hongkui, Shoreline, WA, UNITED STATES  
 PATENT ASSIGNEE(S): Nura Inc., Seattle, WA, UNITED STATES, 98104 (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20060134109	A1	20060622
APPLICATION INFO.:	US 2003-527265	A1	20030909 (10)
	WO 2003-US28226		20030909
			20060126 PCT 371 date
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 2002-409303, filed on 9 Sep 2002, PENDING Continuation-in-part of Ser. No. US 2003-461329, filed on 9 Apr 2003, PENDING		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	APPLICATION		
LEGAL REPRESENTATIVE:	SEED INTELLECTUAL PROPERTY LAW GROUP PLLC, 701 FIFTH AVE, SUITE 6300, SEATTLE, WA, 98104-7092, US		
NUMBER OF CLAIMS:	34		
EXEMPLARY CLAIM:	1-642		
NUMBER OF DRAWINGS:	9 Drawing Page(s)		
LINE COUNT:	15018		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM Diseases of the colon that can be treated or diagnosed using the methods of the invention or for which candidate therapeutic compounds may be identified include acute self-limited infectious colitis, adenocarcinoma, adenoma, adenoma-carcinoma sequence, adenomatous polyposis coli, adenosquamous carcinomas, allergic (eosinophilic) proctitis and colitis, amebiasis, amyloidosis, angiodysplasia, anorectal malformations, blue rubber bleb nevus syndrome, brown bowel syndrome, Campylobacter fetus infection, carcinoid tumors, carcinoma of the anal canal, carcinoma of the colon and rectum, chlamidial proctitis, Crohn's disease, clear cell carcinomas, Clostridium difficile pseudomembranous enterocolitis, collagenous colitis, colonic adenoma, colonic diverticulosis, colonic inertia, colonic ischemia, congenital atresia, congenital megacolon (Hirschsprung's disease), congenital stenosis, constipation, Cowden's syndrome, cystic fibrosis, cytomegalovirus colitis, diarrhea, dieulafor lesion, diversion colitis, diverticulitis,

diverticulosis, drug-induced diseases, dysplasia and malignancy in inflammatory bowel disease, Ehlers-Danlos syndromes, enterobiasis, familial adenomatous polyposis, familial polyposis syndromes, Gardner's syndrome, gastrointestinal stromal neoplasms, hemangiomas and vascular anomalies, hemorrhoids, hereditary hemorrhagic telangiectasia, herpes colitis, hyperplastic polyps, idiopathic inflammatory bowel disease, incontinence, inflammatory bowel syndrome, inflammatory polyps, inherited adenomatous polyposis syndromes, intestinal hamartomas, intestinal pseudo-obstruction, irritable bowel syndrome, ischemic colitis, juvenile polyposis, juvenile polyps, Klippel-Trenaunay-Weber syndrome, leiomyomas, lipomas, lymphocytic (microscopic) colitis, lymphoid hyperplasia and lymphoma, malaknock outplakia, malignant lymphoma, malignant neoplasms, malrotation, metastatic neoplasms, mixed hyperplastic and adenomatous polyps, mucosal prolapse syndrome, neonatal necrotizing enterocolitis, neuroendocrine cell tumors, neurogenic tumors, neutropenic enterocolitis, non-neoplastic polyps, Peutz-Jeghers syndrome, pneumatosis cystoides intestinalis, polyposis coli, pseudomembranous colitis, pseudoxanthoma elasticum, pure squamous carcinomas, radiation colitis, schistosomiasis, Shigella colitis (bacillary dysentery), spindle cell carcinomas, spirochetosis, stercoral ulcers, stromal tumors, systemic sclerosis and CREST syndrome, trichuriasis, tubular adenoma (adenomatous polyp, polypoid adenoma), Turcot's syndrome, Turner's syndrome, ulcerative colitis, villous adenoma, and volvulus.

DETD Exemplary diseases and disorders involving the colon include acute self-limited infectious colitis, adenocarcinoma, adenoma, adenoma-carcinoma sequence, adenomatous polyposis coli, adenosquamous carcinomas, allergic (eosinophilic) proctitis and colitis, amebiasis, amyloidosis, angiodysplasia, anorectal malformations, blue rubber bleb nevus syndrome, brown bowel syndrome, Campylobacter fetus infection, carcinoid tumors, carcinoma of the anal canal, carcinoma of the colon and rectum, chlamydial proctitis, Crohn's disease, clear cell carcinomas, Clostridium difficile pseudomembranous enterocolitis, collagenous colitis, colonic adenoma, colonic diverticulosis, colonic inertia, colonic ischemia, congenital atresia, congenital megacolon (Hirschsprung's disease), congenital stenosis, constipation, Cowden's syndrome, cystic fibrosis, cytomegalovirus colitis, diarrhea, dieulafor lesion, diversion colitis, diverticulitis, diverticulosis, drug-induced diseases, dysplasia and malignancy in inflammatory bowel disease, Ehlers-Danlos syndromes, enterobiasis, familial adenomatous polyposis, familial polyposis syndromes, Gardner's syndrome, gastrointestinal stromal neoplasms, hemangiomas and vascular anomalies, hemorrhoids, hereditary hemorrhagic telangiectasia, herpes colitis, hyperplastic polyps, idiopathic inflammatory bowel disease, incontinence, inflammatory bowel syndrome, inflammatory polyps, inherited adenomatous polyposis syndromes, intestinal hamartomas, intestinal pseudo-obstruction, irritable bowel syndrome, ischemic colitis, juvenile polyposis, juvenile polyps, Klippel-Trenaunay-Weber syndrome, leiomyomas, lipomas, lymphocytic (microscopic) colitis, lymphoid hyperplasia and lymphoma, malaknock outplakia, malignant lymphoma, malignant neoplasms, malrotation, metastatic neoplasms, mixed hyperplastic and adenomatous polyps, mucosal prolapse syndrome, neonatal necrotizing enterocolitis, neuroendocrine cell tumors, neurogenic tumors, neutropenic enterocolitis, non-neoplastic polyps, Peutz-Jeghers syndrome, pneumatosis cystoides intestinalis, polyposis coli, pseudomembranous colitis, pseudoxanthoma elasticum, pure squamous carcinomas, radiation colitis, schistosomiasis, Shigella colitis (bacillary dysentery), spindle cell carcinomas, spirochetosis, stercoral ulcers, stromal tumors, systemic sclerosis and CREST syndrome, trichuriasis, tubular adenoma (adenomatous polyp, polypoid adenoma),

Turcot's syndrome, Turner's syndrome, ulcerative colitis, villous adenoma, and volvulus.

DETD The results of RT-PCR analysis with 100 different GPCRs and 26 mouse tissues (17 peripheral tissues and 9 brain regions) are shown in FIG. 4. The data is presented as a semi-quantitative scattergram. The most remarkable finding was that 94% of GPCRs were detected in the brain, generally in 4 to 5 distinct anatomical areas. The largest number of genes was detected in the hypothalamus (82 genes), a brain region of high structural complexity. Individual peripheral tissues also showed expression of multiple different GPCRs, ranging from 12 genes in muscle to 69 genes in ovary.

L9 ANSWER 17 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2006:40188 USPATFULL <<LOGINID::20090129>>

TITLE: Immunoglobulin variants and uses thereof

INVENTOR(S): Adams, Camellia W., San Jose, CA, UNITED STATES  
Chan, Andrew C., Menlo Park, CA, UNITED STATES  
Crowley, Craig W., Portola Valley, CA, UNITED STATES  
Lowman, Henry B., El Granada, CA, UNITED STATES  
Nakamura, Gerald R., San Francisco, CA, UNITED STATES  
Presta, Leonard G., San Francisco, CA, UNITED STATES  
PATENT ASSIGNEE(S): Genentech, Inc., South San Francisco, CA, UNITED STATES  
(U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20060034835	A1	20060216
APPLICATION INFO.:	US 2005-147780	A1	20050607 (11)
RELATED APPLN. INFO.:	Continuation of Ser. No. WO 2003-US40426, filed on 16 Dec 2003, PENDING		

	NUMBER	DATE
PRIORITY INFORMATION:	US 2002-434115P	20021216 (60)
	US 2003-526163P	20031201 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	GENENTECH, INC., 1 DNA WAY, SOUTH SAN FRANCISCO, CA, 94080, US	
NUMBER OF CLAIMS:	81	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	25 Drawing Page(s)	
LINE COUNT:	5481	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Expression and cloning vectors may contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g., the gene encoding D-alanine racemase for Bacilli.

DETD Suitable host cells for cloning or expressing the DNA in the vectors herein are the prokaryote, yeast, or higher eukaryote cells described above. Suitable prokaryotes for this purpose include eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as Escherichia, e.g., E. coli, Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. licheniformis (e.g. B. licheniformis 41P disclosed in DD 266,710 published 12 Apr. 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. One preferred E.

coli cloning host is E. coli 294 (ATCC 31,446), although other strains such as E. coli B, E. coli X1776 (ATCC 31,537), and E. coli W3110 (ATCC 27,325) are suitable. These examples are illustrative rather than limiting.

DETD Peripheral B-cell concentrations were determined by a FACS method that counted CD3-/CD40+cells. The percent of CD3-CD40+B cells of total lymphocytes in monkey samples were obtained by the following gating strategy. The lymphocyte population was marked on the forward scatter/side scatter scattergram to define Region 1 (R1). Using events in R1, fluorescence intensity dot plots were displayed for CD40 and CD3 markers. Fluorescently labeled isotype controls were used to determine respective cutoff points for CD40 and CD3 positivity.

L9 ANSWER 18 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2006:27472 USPATFULL <<LOGINID::20090129>>

TITLE: Immunoglobulin variants and uses thereof

INVENTOR(S): Adams, Camellia W., San Jose, CA, UNITED STATES  
Chan, Andrew C., Menlo Park, CA, UNITED STATES  
Crowley, Craig W., Portola Valley, CA, UNITED STATES  
Lowman, Henry B., El Granada, CA, UNITED STATES  
Nakamura, Gerald R., San Francisco, CA, UNITED STATES  
Presta, Leonard G., San Francisco, CA, UNITED STATES  
PATENT ASSIGNEE(S): Genentech, Inc., South San Francisco, CA, UNITED STATES  
(U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20060024300	A1	20060202
APPLICATION INFO.:	US 2005-190364	A1	20050726 (11)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 2005-147780, filed on 7 Jun 2005, PENDING Continuation of Ser. No. WO 2003-US40426, filed on 16 Dec 2003, PENDING		

	NUMBER	DATE
PRIORITY INFORMATION:	US 2003-526163P	20031201 (60)
	US 2002-434115P	20021216 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	GENENTECH, INC., 1 DNA WAY, SOUTH SAN FRANCISCO, CA, 94080, US	
NUMBER OF CLAIMS:	13	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	29 Drawing Page(s)	
LINE COUNT:	5333	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Expression and cloning vectors may contain a selection gene, also termed a selectable marker. Typical selection genes encode proteins that (a) confer resistance to antibiotics or other toxins, e.g., ampicillin, neomycin, methotrexate, or tetracycline, (b) complement auxotrophic deficiencies, or (c) supply critical nutrients not available from complex media, e.g., the gene encoding D-alanine racemase for Bacilli.

DETD Suitable host cells for cloning or expressing the DNA in the vectors herein are the prokaryote, yeast, or higher eukaryote cells described above. Suitable prokaryotes for this purpose include eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as Escherichia, e.g., E. coli, Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. licheniformis (e.g., B.



licheniformis 41P disclosed in DD 266,710 published 12 Apr. 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. One preferred E. coli cloning host is E. coli 294 (ATCC 31,446), although other strains such as E. coli B, E. coli X1776 (ATCC 31,537), and E. coli W31 10 (ATCC 27,325) are suitable. These examples are illustrative rather than limiting.

DETD Peripheral B-cell concentrations were determined by a FACS method that counted CD3-/CD40+ cells. The percent of CD3-CD40+ B cells of total lymphocytes in monkey samples were obtained by the following gating strategy. The lymphocyte population was marked on the forward scatter/side scatter scattergram to define Region 1 (R1). Using events in R1, fluorescence intensity dot plots were displayed for CD40 and CD3 markers. Fluorescently labeled isotype controls were used to determine respective cutoff points for CD40 and CD3 positivity.

L9 ANSWER 19 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2005:188867 USPATFULL <<LOGINID::20090129>>

TITLE: Combination therapy for B cell disorders

INVENTOR(S): Chan, Andrew, Menlo Park, CA, UNITED STATES

Gong, Qian, Foster City, CA, UNITED STATES

Martin, Flavius, Hayward, CA, UNITED STATES

PATENT ASSIGNEE(S): Genentech, Inc., South San Francisco, CA, UNITED STATES (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20050163775	A1	20050728
APPLICATION INFO.:	US 2004-21874	A1	20041222 (11)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 2004-861049, filed on 4 Jun 2004, PENDING		

	NUMBER	DATE
PRIORITY INFORMATION:	US 2003-476531P	20030606 (60)
	US 2003-476414P	20030605 (60)
	US 2003-476481P	20030605 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	MERCHANT & GOULD PC, P.O. BOX 2903, MINNEAPOLIS, MN, 55402-0903, US	
NUMBER OF CLAIMS:	58	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	37 Drawing Page(s)	
LINE COUNT:	6855	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes for this purpose include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as Escherichia, e.g., E. coli, Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. licheniformis (e.g., B. licheniformis 41P disclosed in DD 266,710 published 12 Apr. 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. Preferably, the host cell should secrete minimal amounts of proteolytic enzymes.

DETD Peripheral B-cell concentrations are determined by a FACS method that count CD3-/CD40+ cells. The percent of CD3-CD40+ B cells of total lymphocytes in samples can be obtained by the following gating strategy. The lymphocyte population is marked on the forward scatter/side scatter scattergram to define Region 1 (R1). Using events in R1,

fluorescence intensity dot plots are displayed for CD40 and CD3 markers. Fluorescently labeled isotype controls are used to determine respective cutoff points for CD40 and CD3 positivity.

L9 ANSWER 20 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2005:124263 USPATFULL <<LOGINID::20090129>>  
 TITLE: ELECTROCHEMILUMINESCENT ASSAYS  
 INVENTOR(S): Massey, Richard J., Rockville, MD, UNITED STATES  
 Powell, Michael J., Rockville, MD, UNITED STATES  
 Mied, Paul A., New Windsor, MD, UNITED STATES  
 Feng, Peter, Rockville, MD, UNITED STATES  
 Della Ciana, Leopoldo, Rockville, MD, UNITED STATES  
 Dressick, Walter J., Rockville, MD, UNITED STATES  
 Poonian, Mohindar S., Gaithersburg, MD, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20050106652	A1	20050519
	US 6916606	B2	20050712
APPLICATION INFO.:	US 2002-274079	A1	20021018 (10)
RELATED APPLN. INFO.:	Division of Ser. No. US 1995-415756, filed on 3 Apr 1995, ABANDONED Continuation of Ser. No. US 1994-195825, filed on 10 Feb 1994, ABANDONED Continuation of Ser. No. US 1987-369560, filed on 18 Dec 1987, ABANDONED Continuation-in-part of Ser. No. US 1986-858354, filed on 30 Apr 1986, ABANDONED		

	NUMBER	DATE
PRIORITY INFORMATION:	WO 1987-US987	19870430
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	FINNEGAN, HENDERSON, FARABOW, GARRETT & DUNNER, LLP, 901 NEW YORK AVENUE, NW, WASHINGTON, DC, 20001-4413, US	
NUMBER OF CLAIMS:	3	
EXEMPLARY CLAIM:	1-156	
NUMBER OF DRAWINGS:	13 Drawing Page(s)	
LINE COUNT:	3991	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD The analytes of interest may be microorganisms. The microorganisms may be viable or nonviable. Additionally, the microorganisms may be bacteria. Examples of bacteria which may detected by this method include, but are not limited to, Salmonella, Campylobacter, Escherichia, Yersinia, Bacillus, Vibrio, Legionella, Clostridium, Streptococcus or Staphylococcus.

DETD Each serum sample was also analyzed for the concentration of theophylline by a fluorescence polarization assay. The concentration of theophylline measured by the homogeneous electrochemiluminescence immunoassay and the fluorescence polarization assay were compared. The data were plotted as a scattergram and are shown in FIGS. 4A-D. The data points were analyzed by linear regression and the correlation coefficients were calculated. The analysis demonstrates an excellent correlation between the two assays. The correlation coefficients (r) were between 0.98 and 1.00. The slopes of the curves for normal, hemolyzed, and lipemic serum samples were between 0.8 and 1.2, demonstrating excellent recovery of theophylline from these serum samples.

DETD The results for the homogeneous electrochemiluminescent immunoassay and the HPLC assay for determining the concentration of theophylline in serum are shown in FIG. 5. The data were plotted as a scattergram and the data points were analyzed by linear

regression. The correlation coefficient was calculated. The correlation coefficient (r) was 0.98, which demonstrates excellent correlation between the two assays.

L9 ANSWER 21 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2005:111165 USPATFULL <<LOGINID::20090129>>  
TITLE: Combination therapy for B cell disorders  
INVENTOR(S): Chan, Andrew, Menlo Park, CA, UNITED STATES  
Gong, Qian, Foster City, CA, UNITED STATES  
Martin, Flavius, Hayward, CA, UNITED STATES  
PATENT ASSIGNEE(S): Genentech, Inc., South San Francisco, CA, UNITED STATES, 94080 (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20050095243	A1	20050505
APPLICATION INFO.:	US 2004-861049	A1	20040604 (10)

	NUMBER	DATE
PRIORITY INFORMATION:	US 2003-476531P	20030606 (60)
	US 2003-476414P	20030605 (60)
	US 2003-476481P	20030605 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	GENENTECH, INC., 1 DNA WAY, SOUTH SAN FRANCISCO, CA, 94080, US	
NUMBER OF CLAIMS:	55	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	36 Drawing Page(s)	
LINE COUNT:	4451	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Suitable host cells for cloning or expressing the DNA in the vectors herein include prokaryote, yeast, or higher eukaryote cells. Suitable prokaryotes for this purpose include but are not limited to eubacteria, such as Gram-negative or Gram-positive organisms, for example, Enterobacteriaceae such as Escherichia, e.g., E. coli, Enterobacter, Erwinia, Klebsiella, Proteus, Salmonella, e.g., Salmonella typhimurium, Serratia, e.g., Serratia marcescans, and Shigella, as well as Bacilli such as B. subtilis and B. licheniformis (e.g., B. licheniformis 41P disclosed in DD 266,710 published 12 Apr. 1989), Pseudomonas such as P. aeruginosa, and Streptomyces. Preferably, the host cell should secrete minimal amounts of proteolytic enzymes.

DETD Peripheral B-cell concentrations are determined by a FACS method that count CD3-/CD40+ cells. The percent of CD3-CD40+ B cells of total lymphocytes in samples can be obtained by the following gating strategy. The lymphocyte population is marked on the forward scatter/ side scatter scattergram to define Region 1 (R1). Using events in R1, fluorescence intensity dot plots are displayed for CD40 and CD3 markers. Fluorescently labeled isotype controls are used to determine respective cutoff points for CD40 and CD3 positivity.

L9 ANSWER 22 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2004:227395 USPATFULL <<LOGINID::20090129>>  
TITLE: Method of staining, detecting and counting bacteria, and a diluent for bacterial stain  
INVENTOR(S): Sakai, Yasuhiro, Hyogo, JAPAN  
Kawashima, Yasuyuki, Hyogo, JAPAN  
Inoue, Junya, Hyogo, JAPAN  
Ikeuchi, Yoshiro, Hyogo, JAPAN  
PATENT ASSIGNEE(S): Sysmex Corporation (non-U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20040175781	A1	20040909
APPLICATION INFO.:	US 2004-803667	A1	20040318 (10)
RELATED APPLN. INFO.:	Division of Ser. No. US 2001-5753, filed on 29 Oct 2001, PENDING		

	NUMBER	DATE
PRIORITY INFORMATION:	JP 2000-334641	20001101
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	Lance J. Lieberman, Esq., Cohen, Pontani, Lieberman & Pavane, Suite 1210, 551 Fifth Avenue, New York, NY, 10176	
NUMBER OF CLAIMS:	23	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	2 Drawing Page(s)	
LINE COUNT:	558	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM [0009] The microscopic examination of bacteria without staining treatment can be carried out quickly, but it cannot discriminate bacteria particularly when coccus contaminants are contained.

DRWD [0022] FIG. 1 is a scattergram of a fluorescent light intensity--a forward scattered light intensity obtained in the case where ascorbic acid is used as a reducing agent in Example 1 of the present invention;

DRWD [0023] FIG. 2 is a scattergram of a fluorescent light intensity--a forward scattered light intensity obtained in the case where the reducing agent is not used in Example 1 of the present invention;

DRWD [0024] FIG. 3 is a scattergram of a fluorescent light intensity--a forward scattered light intensity obtained in the case where sulfamic acid is used as the reducing agent in Example 2 of the present invention; and

DETD [0026] In the present invention, the sample is not particularly limited as long as it is a sample to be examined for the presence or absence of bacteria and to count a number of bacteria if the sample contains bacteria. Bacteria referred herein include bacteria which reduce nitrite and produce nitrous acid, e.g., intestinal bacteria such as *Staphyrococcus aureus*, Gram-negative facultative bacilli such as *E. coli*, *Klebsiella* sp. and *Proteus* sp., or bacteria observed in a urine sample such as *E. coli*, *Klebsiella* sp., as well as *Staphyrococcus* sp., *Pseudomonas* sp., *Serratia* sp., *Enterobacter* sp., *Enterococcus* sp., *Streptococcus* sp. and *Citrobacter* sp. For example, the sample may be a clinical sample such as urine, blood, spinal fluid or the like. The sample may be diluted with purified water or the like two or more times, preferably 4 to 15 times, more preferably 5 to 10 times. The present invention is particularly effective for a urine sample.

DETD [0069] Discrimination of bacteria from other components and counting of bacteria are carried out in accordance with combination of signals obtained by using a flow cytometer. Example of the combination includes, for example, a forward scattered light intensity and a forward scattered light pulse width, a forward scattered light intensity and a fluorescent light intensity, a forward scattered light pulse width and a fluorescent light intensity, and the like. In a suitable manner, for example, firstly, a scattergram is formed from the combination of the forward scattered light intensity and the forward scattered light pulse width, and then gating is performed to a mass including bacteria

specified on the scattergram to separate mucus threads, mainly. Further, another scattergram is formed from the forward scattered light intensity and the fluorescent light intensity of the gated mass to-separate bacteria from other components (crystals, cell fragments and the like) based on the difference in the fluorescent light intensity. The outline of the method is shown in FIG. 4. Where the sample contains bacteria only, a scattergram is formed from the forward scattered light intensity and the fluorescent light intensity to count them.

DETD [0075] To 140  $\mu$ l of a sample containing a large amount of nitrite ions (bacteria concentration of  $5.0 \times 10^6$ /ml; hospital urine), 952  $\mu$ l of the above-mentioned diluent was added and the staining solution was added so that the final concentration of the dye A would be 1 ppm. Staining was carried out at 40° C. for 20 seconds and then scattered light and fluorescent light were measured by a flow cytometer provided with a red semiconductor laser as a light source (amount of examined urine: 8.0  $\mu$ l). Then, as shown in FIG. 1, a scattergram was formed with a fluorescent light intensity (FLI) as an horizontal axis and a forward scattered light intensity (FSLI) as a vertical axis. As a control, measurement was performed using a reagent containing no ascorbic acid (FIG. 2).

L9 ANSWER 23 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2004:203888 USPATFULL <<LOGINID::20090129>>  
 TITLE: CTL inducing peptides from c-erb2 (HER-2/neu)  
 INVENTOR(S): Grey, Howard M., La Jolla, CA, UNITED STATES  
 Sette, Alessandro, La Jolla, CA, UNITED STATES  
 Sidney, John, La Jolla, CA, UNITED STATES  
 PATENT ASSIGNEE(S): Epimmune Inc. (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20040157780	A1	20040812
APPLICATION INFO.:	US 2004-770493	A1	20040204 (10)
RELATED APPLN. INFO.:	Continuation of Ser. No. US 1994-205713, filed on 4 Mar 1994, PENDING Continuation-in-part of Ser. No. US 1993-159184, filed on 29 Nov 1993, ABANDONED		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	APPLICATION		
LEGAL REPRESENTATIVE:	STERNE, KESSLER, GOLDSTEIN & FOX PLLC, 1100 NEW YORK AVENUE, N.W., WASHINGTON, DC, 20005		
NUMBER OF CLAIMS:	23		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	5 Drawing Page(s)		
LINE COUNT:	2850		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DRWD [0021] FIG. 2 shows a scattergram of the log of relative binding plotted against the "Grouped Ratio" algorithm for 9 mer peptides.

DRWD [0022] FIG. 3 shows a scattergram of the log of relative binding plotted against the average "Log of Binding" algorithm score for 9 mer peptides.

DRWD [0023] FIGS. 4 and 5 show scattergrams of a set of 10-mer peptides containing preferred residues in positions 2 and 10 as scored by the "Grouped Ratio" and "Log of Binding" algorithms.

DETD [0079] For therapeutic or immunization purposes, the peptides of the invention can also be expressed by attenuated viral hosts, such as vaccinia or fowlpox. This approach involves the use of vaccinia virus as a vector to express nucleotide sequences that encode the peptides of the invention. Upon introduction into an acutely or chronically infected host or into a non-infected host, the recombinant vaccinia virus



F	3.00	0.01	2.60	3.10	3.60	0.60	1.60	14.1	2.10
G	0.80	0.01	0.50	4.70	0.80	6.30	2.70	0.70	0.80
H	1.20	0.01	0.30	0.10	0.70	0.40	0.20	0.01	0.20
I	3.00	0.50	10.2	1.00	1.30	2.10	1.40	4.70	0.80
K	1.20	0.01	0.30	0.10	0.70	0.40	0.20	0.01	0.20
L	3.00	1.10	10.2	1.00	1.30	2.10	1.40	4.70	0.80
M	3.00	0.60	10.2	1.00	1.30	2.10	1.40	4.70	0.80
N	1.00	0.01	0.90	0.80	0.80	0.80	0.60	0.40	0.70
P	0.00	0.01	0.40	2.60	0.01	1.00	0.40	1.90	1.20
Q	1.00	0.01	0.90	0.80	0.80	0.80	0.60	0.40	0.70
R	1.20	0.01	0.30	0.10	0.70	0.40	0.20	0.01	0.20
S	0.90	0.01	0.90	1.10	1.00	0.90	1.40	1.30	2.90
T	0.90	0.01	0.90	1.10	1.00	0.90	1.40	1.30	2.90
V	3.00	0.10	10.2	1.00	1.30	2.10	1.40	4.70	0.80
W	3.00	0.01	2.60	3.10	3.60	0.60	1.60	14.1	2.10
Y	3.00	0.01	2.60	3.10	3.60	0.60	1.60	14.1	2.10

L9 ANSWER 24 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2003:288180 USPATFULL <<LOGINID::20090129>>  
 TITLE: Hematopoietic growth factor inducible neurokinin-1 gene  
 INVENTOR(S): Rameshwar, Pranela, Maplewood, NJ, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20030202938	A1	20031030
APPLICATION INFO.:	US 2003-463106	A1	20030617 (10)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 2001-39272, filed on 20 Oct 2001, PENDING		

	NUMBER	DATE
PRIORITY INFORMATION:	US 2000-241881P	20001020 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	PERKINS COIE LLP, POST OFFICE BOX 1208, SEATTLE, WA, 98111-1208	
NUMBER OF CLAIMS:	20	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	16 Drawing Page(s)	
LINE COUNT:	3549	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD [0175] Representative examples of appropriate hosts for in vitro procedures include bacterial cells, such as streptococci, staphylococci, E. coli, Streptomyces and Bacillus subtilis cells; fungal cells, such as yeast cells and Aspergiffits cells, insect cells such as

Drosophila S2 and Spodoptera Sf9 cells; animal cells such as CHO, COS, HeLa, C127, 3T3, BHK, HEK 293 and Bowes melanoma cells, and plant cells. The selection of an appropriate host is deemed to be within the scope of those skilled in the art from the teachings herein.

DETD [0294] Another experiment of the present invention characterizes slow-growing and/or drug resistant clones by flow cytometry, which determines the degree that cells from different clones can pump dye (either Rhodamine 123 or Hoechst as used experimentally). The cancer stem cells are likely more efficient than cancer progenitors to pump dye out of cells. The size and scatter pattern of the different clones are examined to determine whether the slow-growing clones represent side population (S-Pop) cells and whether the progenitor cells larger so that they would be identified at a particular region in the scattergram. A subset of the study population is collected by cell sorting based on size and/or rhodamine uptake. Drug resistant cells are categorized as S-Pop, S-Pop/Rhodamine or Hoescht.sup.dim, S-Pop/Rhodamine or Hoescht.sup.bright, Forward scatter (FSc), FSc/Rhodamine or Hoescht.sup.dim;, FSc/Rhodamine or Hoesch.sup.bright.

L9 ANSWER 25 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2003:264808 USPATFULL <<LOGINID::20090129>>  
TITLE: HLA-A2.1 binding peptides and their uses  
INVENTOR(S): Grey, Howard M., La Jolla, CA, UNITED STATES  
Sette, Alessandro, La Jolla, CA, UNITED STATES  
Sidney, John, La Jolla, CA, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20030185822	A1	20031002
APPLICATION INFO.:	US 2002-116557	A1	20020403 (10)
RELATED APPLN. INFO.:	Continuation of Ser. No. US 1994-349177, filed on 2 Dec 1994, PENDING Continuation-in-part of Ser. No. US 1993-159184, filed on 29 Nov 1993, ABANDONED Continuation-in-part of Ser. No. US 1993-73205, filed on 4 Jun 1993, ABANDONED Continuation-in-part of Ser. No. US 1993-27146, filed on 5 Mar 1993, ABANDONED		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	APPLICATION		
LEGAL REPRESENTATIVE:	MORRISON & FOERSTER LLP, 3811 VALLEY CENTRE DRIVE, SUITE 500, SAN DIEGO, CA, 92130-2332		
NUMBER OF CLAIMS:	18		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	5 Drawing Page(s)		
LINE COUNT:	3171		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM [0068] For therapeutic or immunization purposes, the peptides of the invention can also be expressed by attenuated viral hosts, such as vaccinia or fowlpox. This approach involves the use of vaccinia virus as a vector to express nucleotide sequences that encode the peptides of the invention. Upon introduction into an acutely or chronically infected host or into a non-infected host, the recombinant vaccinia virus expresses the immunogenic peptide, and thereby elicits a host CTL response. Vaccinia vectors and methods useful in immunization protocols are described in, e.g., U.S. Pat. No. 4,722,848, incorporated herein by reference. Another vector is BCG (Bacille Calmette Guerin). BCG vectors are described in Stover et al. (Nature 351:456-460 (1991)) which is incorporated herein by reference. A wide variety of other vectors useful for therapeutic administration or immunization of the peptides of the invention, e.g., Salmonella typhi vectors and the like, will be apparent to those skilled in the art from the description herein.



DETD [0134] In the present "Grouped Ratio" algorithm residues have been grouped by similarity. This avoids the problem encountered with some rare residues, such as tryptophan, where there are too few occurrences to obtain a statistically significant ratio. Table 16 is a listing of scores obtained by grouping for each of the twenty amino acids by position for 9-mer peptides containing perfect 2/9 motifs. A peptide is scored in the "Grouped Ratio" algorithm as a product of the scores of each of its residues. In the case of positions other than 2 and 9, the scores have been derived using a set of peptides which contain only preferred residues in positions 2 and 9. To enable us to extend our "Grouped Ratio" algorithm to peptides which may have residues other than the preferred ones at 2 and 9, scores for 2 and 9 have been derived from a set of peptides which are single amino acid substitutions at positions 2 and 9. FIG. 2 shows a scattergram of the log of relative binding plotted against "Grouped Ratio" algorithm score for our collection of 9-mer peptides from the previous example.

TABLE 16

	1	2	3	4	5	6	7	8	9
A	2.6	0.03	0.87	0.87	0.65	0.87	4.4	0.29	0.16
C	0.73	0.01	1.9	4.8	0.87	1.2	1.2	1.1	0.01
D	0.10	0.01	0.10	0.65	0.29	0.65	0.11	0.87	0.01
E	0.10	0.01	0.10	0.65	0.29	0.65	0.11	0.87	0.01
F	7.0	0.01	5.2	0.87	8.7	2.0	2.3	2.6	0.01
G	3.5	0.01	0.44	1.1	1.1	1.3	0.44	0.44	0.01
H	3.1	0.01	0.22	1.0	0.87	0.09	0.10	1.3	0.01
I	3.1	0.14	1.8	0.55	0.87	1.4	1.2	1.8	0.40
K	3.1	0.01	0.22	1.0	0.87	0.09	0.10	1.3	0.01
L	3.1	1.00	1.8	0.55	0.87	1.4	1.2	1.8	0.09
M	3.1	2.00	1.8	0.55	0.87	1.4	1.2	1.8	0.06
N	0.50	0.01	0.37	1.2	0.87	1.1	0.65	0.33	0.01
P	0.12	0.01	0.70	0.73	2.6	1.8	2.9	0.10	0.01
Q	0.50	0.01	0.37	1.2	0.87	1.1	0.65	0.33	0.01
R	3.1	0.01	0.22	1.0	0.87	0.09	0.10	1.3	0.01
S	0.73	0.01	1.9	4.8	0.87	1.2	1.2	1.1	0.01
T	0.73	0.01	1.9	4.8	0.87	1.2	1.2	1.1	0.01
V	3.1	0.08	1.8	0.55	0.87	1.4	1.2	1.8	1.00
W	7.0	0.01	5.2	0.87	8.7	2.0	2.3	2.6	0.01
Y	7.0	0.01	5.2	0.87	8.7	2.0	2.3	2.6	0.01

DETD [0137] An algorithm using the average binding affinity of all the peptides with a certain amino acid (or amino acid type) at a certain position has the advantage of including all of the peptides in the analysis, and not just good/intermediate binders and non-binders. Moreover, it gives a more quantitative measure of affinity than the simpler "Grouped Ratio" algorithm. We have created such an algorithm by calculating for each amino acid, by position, the average log of binding when that particular residue occurs in our set of 161 2,9 motif containing peptides. These values are shown in Table 18. The algorithm score for a peptide is then taken as the sum of the scores by position for each residues. FIG. 3 shows a scattergram of the log of relative binding against the average "Log of Binding" algorithm score. Table 17 shows the ability of the two algorithms to predict peptide binding at various levels, as a function of the cut-off score used. The ability of a 2,9 motif to predict binding in the same peptide set is also shown for reference purposes. It is clear from this comparison that both algorithms of this invention have a greater ability to predict populations with higher frequencies of good binders than a 2,9 motif

alone. Differences between the "Grouped Ratio" algorithm and the "Log of Binding" algorithm are small in the set of peptides analyzed here, but do suggest that the "Log of Binding" algorithm is a better, if only slightly, predictor than the "Grouped Ratio" algorithm.

DETD [0138] Using the methods described in the proceeding example, an analogous set of algorithms has been developed for predicting the binding of 10-mer peptides. Table 19 shows the scores used in a "Grouped Ratio" algorithm, and Table 20 shows the "Log of Binding" algorithm scores, for 10-mer peptides. Table 21 shows a comparison of the application of different algorithmic methods to select binding peptides. FIGS. 4 and 5 show, respectively, scattergrams of a set of 10-mer peptides containing preferred residues in positions 2 and 10 as scored by the "Grouped Ratio" and "Log of Binding" algorithms.

L9 ANSWER 26 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2003:120179 USPATFULL <<LOGINID::20090129>>  
 TITLE: Orthologues of human receptors and methods of use  
 INVENTOR(S): Horlick, Robert, San Diego, CA, UNITED STATES  
 Zhao, Jiuquao, Hockessin, DE, UNITED STATES  
 Swanson, Robert, Cranbury, NJ, UNITED STATES  
 Webb, Maria, Flemington, NJ, UNITED STATES  
 Strohl, Barbara, Hamilton, NJ, UNITED STATES  
 Baldwin, John J., Gwynedd Valley, PA, UNITED STATES  
 Auld, Douglas S., Cranbury, NJ, UNITED STATES  
 Chen, Xiao Ge, Princeton, NJ, UNITED STATES

	NUMBER	KIND	DATE
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PATENT INFORMATION:	US 20030082660	A1	20030501
	US 7041463	B2	20060509
APPLICATION INFO.:	US 2002-237563	A1	20020909 (10)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 2000-576160, filed on 22 May 2000, GRANTED, Pat. No. US 6469150		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	APPLICATION		
LEGAL REPRESENTATIVE:	HESLIN ROTHENBERG FARLEY & MESITI PC, 5 COLUMBIA CIRCLE, ALBANY, NY, 12203		
NUMBER OF CLAIMS:	55		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	19 Drawing Page(s)		
LINE COUNT:	1224		
CAS INDEXING IS AVAILABLE FOR THIS PATENT.			

DETD [0063] Representative examples of appropriate hosts include bacterial cells, such as E. coli, Streptomyces and Bacillus subtilis cells; fungal cells, such as yeast cells and Aspergillus cells; insect cells such as Drosophila S2 and Spodoptera Sf9 cells; animal cells and plant cells. One of skill in the art will recognize that different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins and gene products. Host cells suitable for expression of the inserted receptor sequences of the present invention are those having the capability to effect such post-translational modifications as necessary to produce a functional receptor. Suitable mammalian host cells include but are not limited to CHO, VERO, BHK, HeLa, COS, MDCK, HEK 293, 3T3, W138.

DETD [0094] A subset of compounds that retain significant potency at most of the receptors was identified. Comparison of the activity of the compounds at animal vs. human B.sub.1 receptor orthologues is shown in the scattergrams of FIGS. 4-6. The solid line at 45° in each panel represents an isocline of equal potency. FIGS. 4-6 show two independent human data sets compared to each other, and species-to-human comparisons, as labeled at the top of each scattergram. The

correlation coefficients of each pair of data sets is shown below each figure. Compounds were tested at 1  $\mu$ M concentration at the rat B.sub.1 receptor, and at 0.1  $\mu$ M concentration at all other animal orthologues. Displacement was tested in the presence of 1.5 nM [<sup>3</sup>H]-dAKd for rat B.sub.1, and 0.6 nM for all other B.sub.1 receptors. Data points are marked as follow: .quadrature., PS978163; .diamond., PS596668; .largecircle., PS972282;  $\Delta$ , PS309799. Conversely, a subset of compounds that exhibit considerable differences in specificity among the orthologues was also identified (data point for PS309799 shown enclosed by triangle). To verify the validity of the scattergram results, the potencies of these four non-peptide compounds were further assessed by ligand displacement assays at the B.sub.1 orthologous receptors. A comparison of K.sub.1s among the four compounds revealed dramatic differences in species specificity. Compound PS309799 showed the greatest variation of activity, ranging from low nM potency in tree shrew and human to inactive at dog and rabbit. PS596668 had a similar activity profile to PS309799 except it demonstrated potent activity at the rabbit B1. The remaining two compounds, PS972282 and PS978163, had measurable affinity constants at all six species, although PS978163 was considerably weaker at pig and dog.

L9 ANSWER 27 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2002:300808 USPATFULL <<LOGINID::20090129>>

TITLE: Fusion cells and cytokine compositions for treatment of disease

INVENTOR(S): Ohno, Tsuneya, Boston, MA, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20020168351	A1	20021114
APPLICATION INFO.:	US 2001-12134	A1	20011022 (10)

	NUMBER	DATE
PRIORITY INFORMATION:	US 2000-242154P	20001020 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	PENNIE AND EDMONDS, 1155 AVENUE OF THE AMERICAS, NEW YORK, NY, 100362711	
NUMBER OF CLAIMS:	30	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	15 Drawing Page(s)	
LINE COUNT:	2136	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DRWD [0049] FIG. 8. FACS analysis, cells stained with both PKH-2GL and PKH-26, which were considered to be fusions of DCs and BNL cells, are shown in upper area of cell scattergram with high forward scatter and high side scatter. The cell fraction of high and moderate forward scatter and low side scatter contained many non-fused BNL cells, which those of low forward scatter and low side scatter contained non-fused DCs and non-fused BNL cells. About 30% of the nonadherent cells were fusions as judged from the width of area of double positive cells occupying in the whole scattergram.

DRWD [0050] FIG. 9. FACS analysis of the cell fractions positive for both PKH-2GL and PKH-26 gated on scattergram and examined for antigen expression. I-A.sup.d/I-E.sup.d (MCH class II), CD80, CD86 and CD54 molecules, which are found on DCs, were expressed by the fusions

DETD [0140] In another embodiment, bacterial infections can be treated or prevented such as, but not limited to disorders caused by pathogenic bacteria including, but not limited to, Streptococcus pyogenes, Streptococcus pneumoniae, Neisseria gonorrhoea, Neisseria meningitidis,

Corynebacterium diphtheriae, Clostridium botulinum, Clostridium perfringens, Clostridium tetani, Haemophilus influenzae, Klebsiella pneumoniae, Klebsiella ozaenae, Klebsiella rhinoscleromatis, Staphylococcus aureus, Vibrio cholerae, Escherichia coli, Pseudomonas aeruginosa, Campylobacter (Vibrio) fetus, Campylobacter jejuni, Aeromonas hydrophila, Bacillus cereus, Edwardsiella tarda, Yersinia enterocolitica, Yersinia pestis, Yersinia pseudotuberculosis, Shigella dysenteriae, Shigella flexneri, Shigella sonnei, Salmonella typhimurium, Salmonella typhi, Treponema pallidum, Treponema pertense, Treponema carateum, Borrelia vincentii, Borrelia burgdorferi, Leptospira icterohemorrhagiae, Mycobacterium tuberculosis, Toxoplasma gondii, Pneumocystis carinii, Francisella tularensis, Brucella abortus, Brucella suis, Brucella melitensis, Mycoplasma spp., Rickettsia prowazekii, Rickettsia tsutsugumushi, Chlamydia spp., and Helicobacter pylori.

DETD [0211] Prior to PEG treatment, DCs were treated with an FITC conjugated antibody against CD11c and BNL cells were stained with PKH-26. The cells were fused by PEG treatment and observed under a fluorescence microscope. Cells stained with both FITC (green) and PKH-26 (red) were observed among the PEG-treated cells (FIG. 7). For determination of the fusion efficacy, DCs and BNL cells were stained with fluorescent dyes, PKH-2GL and PKH-26, respectively, and then treated with PEG. By FACS analysis, cells stained with both PKH-2GL and PKH-26, which were considered to be fusions of DCs and BNL cells, are shown in upper area of cell scattergram with high forward scatter and high side scatter (FIG. 8). The cell fraction of high and moderate forward scatter and low side scatter contained many non-fused BNL cells, while those of low forward scatter and low side scatter contained non-fused DCs and non-fused BNL cells (FIG. 8). About 30% of the nonadherent cells were fusions as judged from the width of area of double positive cells occupying in the whole scattergram.

DETD [0212] Phenotypes of the fusions were analyzed by FACS. The cell fraction positive for both PKH-2GL and PKH-26 were gated on scattergram and examined for antigen expression. I-A.sup.d/I-E.sup.d (MCH class II), CD80, CD86 and CD54 molecules, which are found on DCs, were expressed by the fusions (FIG. 9).

L9 ANSWER 28 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2002:243038 USPATFULL <<LOGINID::20090129>>  
 TITLE: CaR receptor as a mediator of migratory cell chemotaxis and/or chemokinesis  
 INVENTOR(S): Poznansky, Mark C., Charlestown, MA, UNITED STATES  
 Scadden, David T., Weston, MA, UNITED STATES  
 Olszak, Ivona T., Charlestown, MA, UNITED STATES  
 Brown, Edward M., Milton, MA, UNITED STATES

	NUMBER	KIND	DATE
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PATENT INFORMATION:	US 20020132224	A1	20020919
	US 7176243	B2	20070213
APPLICATION INFO.:	US 2001-2854	A1	20011101 (10)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. WO 2000-US15440, filed on 2 Jun 2000, UNKNOWN		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	APPLICATION		
LEGAL REPRESENTATIVE:	WOLF GREENFIELD & SACKS, PC, FEDERAL RESERVE PLAZA, 600 ATLANTIC AVENUE, BOSTON, MA, 02210-2211		
NUMBER OF CLAIMS:	84		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	6 Drawing Page(s)		
LINE COUNT:	2510		
CAS INDEXING IS AVAILABLE FOR THIS PATENT.			

DRWD [0030] FIG. 1(a): Scattergram showing CaR positive stain on CD14.sup.+ monocytes (upper panel), and inhibition of anti-CaR antibody binding to CaR by preincubating CD 14.sup.+ monocytes with CaR peptide (lower panel); FIG. 1(b) Graphs showing elevation of CD 14.sup.+ intracellular Ca.sup.++ concentration following elevation in the extracellular Ca.sup.++ concentration or addition of the selective CaR activator NPS R-467 in the extracellular medium.

DETD [0098] Bacteria in general include but are not limited to: P. aeruginosa; Bacillus anthracis; E. coli, Enterocytozoon bieneusi; Klebsiella sp.; Klebsiella pneumoniae; Serratia sp.; Pseudomonas sp.; P. cepacia; Acinetobacter sp.; S. epidermis; E. faecalis; S. pneumoniae; S. aureus; Haemophilus sp.; Haemophilus Influenza; Neisseria Sp.; Neisseria gonorrhoeae; Neisseria meningitis; Helicobacter pylori; Bacteroides sp.; Citrobacter sp.; Branhamella sp.; Salmonella sp.; Salmonella typhi; Shigella sp.; S. pyogenes; Proteus sp.; Clostridium sp.; Erysipelothrix sp.; Listeria sp.; Pasteurella multocida; Streptobacillus sp.; Spirillum sp.; Fusospirocheta sp.; Actinomycetes; Mycoplasma sp.; Chlamydiae sp.; Chlamydia trachomatis; Campylobacter jejuni; Cyclospora cayatanensis; Rickettsia sp.; Spirochaeta, including Treponema pallidum and Borrelia sp.; Legionella sp.; Legionella pneumophila; Mycobacteria sp.; Mycobacterium tuberculosis; Ureaplasma sp.; Streptomyces sp.; Trichomonas sp.; and P. mirabilis, as well as toxins, that include, but are not limited to, Anthrax toxin (EF); Adenylate cyclase toxin; Cholera enterotoxin; E. coli LT toxin; Escherichia coli 0157:H7; Shiga toxin; Botulinum Neurotoxin Type A heavy and light chains; Botulinum Neurotoxin Type B heavy and light chains; Tetanus toxin; Tetanus toxin C fragment; Diphtheria toxin; Pertussis toxin; Parvovirus B19; Staphylococcus enterotoxins; Toxic shock syndrome toxin (TSST-1); Erythrogenic toxin; and Vibrio cholerae 0139.

DETD [0147] FIG. 1a: CD14.sup.+ monocytes stain positively for the CaR, and binding of anti-CaR antibody is inhibitable by preincubation with CaR peptide. Purified peripheral blood CD14.sup.+ monocytes (scattergram) were exposed to anti-CaR antibody (solid area in histogram) or isotype control (open area) and examined by flow cytometry. Monocytes were also preincubated with CaR peptide prior to staining with anti-CaR antibody (dashed area). Data represent one of ten independent experiments with comparable results.

L9 ANSWER 29 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2002:148601 USPATFULL <<LOGINID::20090129>>  
TITLE: Method of staining, detection and counting bacteria, and a diluent for bacterial stain  
INVENTOR(S): Sakai, Yasuhiro, Hyogo, JAPAN  
Kawashima, Yasuyuki, Hyogo, JAPAN  
Inoue, Junya, Hyogo, JAPAN  
Ikeuchi, Yoshiro, Hyogo, JAPAN  
PATENT ASSIGNEE(S): Sysmex Corporation (non-U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20020076743	A1	20020620
	US 7309581	B2	20071218
APPLICATION INFO.:	US 2001-5753	A1	20011029 (10)

	NUMBER	DATE
PRIORITY INFORMATION:	JP 2000-334641	20001101
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	Lance J. Lieberman, Esq., Cohen, Pontani, Lieberman &	

Pavane, 551 Fifth Avenue, Suite 1210, New York, NY,  
10176

NUMBER OF CLAIMS: 23

EXEMPLARY CLAIM: 1

NUMBER OF DRAWINGS: 2 Drawing Page(s)

LINE COUNT: 565

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM [0009] The microscopic examination of bacteria without staining treatment can be carried out quickly, but it cannot discriminate bacteria particularly when coccus contaminants are contained.

DRWD [0022] FIG. 1 is a scattergram of a fluorescent light intensity --a forward scattered light intensity obtained in the case where ascorbic acid is used as a reducing agent in Example 1 of the present invention;

DRWD [0023] FIG. 2 is a scattergram of a fluorescent light intensity --a forward scattered light intensity obtained in the case where the reducing agent is not used in Example 1 of the present invention;

DRWD [0024] FIG. 3 is a scattergram of a fluorescent light intensity --a forward scattered light intensity obtained in the case where sulfamic acid is used as the reducing agent in Example 2 of the present invention; and

DETD [0026] In the present invention, the sample is not particularly limited as long as it is a sample to be examined for the presence or absence of bacteria and to count a number of bacteria if the sample contains bacteria. Bacteria referred herein include bacteria which reduce nitrite and produce nitrous acid, e.g., intestinal bacteria such as Staphyrococcus aureus, Gram-negative facultative bacilli such as E. coli, Klebsiella sp. and Proteus sp., or bacteria observed in a urine sample such as E. coli, Klebsiella sp., as well as Staphyrococcus sp., Pseudomonas sp., Serratia sp., Enterobacter sp., Enterococcus sp., Streptococcus sp. and Citrobacter sp. For example, the sample may be a clinical sample such as urine, blood, spinal fluid or the like. The sample may be diluted with purified water or the like two or more times, preferably 4 to 15 times, more preferably 5 to 10 times. The present invention is particularly effective for a urine sample.

DETD [0072] Discrimination of bacteria from other components and counting of bacteria are carried out in accordance with combination of signals obtained by using a flow cytometer. Example of the combination includes, for example, a forward scattered light intensity and a forward scattered light pulse width, a forward scattered light intensity and a fluorescent light intensity, a forward scattered light pulse width and a fluorescent light intensity, and the like. In a suitable manner, for example, firstly, a scattergram is formed from the combination of the forward scattered light intensity and the forward scattered light pulse width, and then gating is performed to a mass including bacteria specified on the scattergram to separate mucus threads, mainly. Further, another scattergram is formed from the forward scattered light intensity and the fluorescent light intensity of the gated mass to separate bacteria from other components (crystals, cell fragments and the like) based on the difference in the fluorescent light intensity. The outline of the method is shown in FIG. 4. Where the sample contains bacteria only, a scattergram is formed from the forward scattered light intensity and the fluorescent light intensity to count them.

DETD [0077] To 140  $\mu$ l of a sample containing a large amount of nitrite ions (bacteria concentration of  $5.0 \times 10^6$ /ml; hospital urine), 952  $\mu$ l of the above-mentioned diluent was added and the staining solution was added so that the final concentration of the dye A would be 1 ppm. Staining was carried out at 40° C. for 20 seconds and then

scattered light and fluorescent light were measured by a flow cytometer provided with a red semiconductor laser as a light source (amount of examined urine: 8.0  $\mu$ l). Then, as shown in FIG. 1, a scattergram was formed with a fluorescent light intensity (FLI) as an horizontal axis and a forward scattered light intensity (FSLI) as a vertical axis. As a control, measurement was performed using a reagent containing no ascorbic acid (FIG. 2).

L9 ANSWER 30 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2002:72649 USPATFULL <<LOGINID::20090129>>  
 TITLE: Liver tissue source  
 INVENTOR(S): Reid, Lola M., Chapel Hill, NC, UNITED STATES  
 Lecluyse, Edward L., Chapel Hill, NC, UNITED STATES

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 20020039786	A1	20020404
APPLICATION INFO.:	US 2001-764359	A1	20010119 (9)

	NUMBER	DATE
PRIORITY INFORMATION:	US 2000-176798P	20000119 (60)
DOCUMENT TYPE:	Utility	
FILE SEGMENT:	APPLICATION	
LEGAL REPRESENTATIVE:	PEPPER HAMILTON, 600 FOURTEENTH STREET NW, WASHINGTON, DC, 20005	
NUMBER OF CLAIMS:	40	
EXEMPLARY CLAIM:	1	
NUMBER OF DRAWINGS:	16 Drawing Page(s)	
LINE COUNT:	2804	

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM [0025] As a further object of the invention a method of therapy is provided, in which progenitor cells are used as a cellular transplant, a bioreactor, an artificial organ, etc. The preferred medical conditions and needs comprise Crigler-Najjar syndrome, tyrosinemia, cirrhosis, acute liver failure, diabetes, and other liver and liver-related conditions known in the art. In general, patients are treated who may suffer from at least one liver disorder selected from the group consisting of inflammation of the liver, viral hepatitis, toxic liver cell damage, fibrosis of the liver, cirrhosis of the liver, liver congestion, liver dystrophy, fatty degeneration of liver cells, fatty liver, disturbances of the detoxification function, disturbances of the excretory function of the liver, disturbances of the conjugational function of the liver, disturbances of the synthesizing function of the liver portal hypertension due to a liver disease, or a liver failure coma, and intoxication by protein degradation products or ammonia. These malfunctions result in diseases such as Alagille syndrome, alcoholic liver disease, alpha-1-antitrypsin deficiency, autoimmune hepatitis, biliary atresia, biliary ductopenia, bone marrow failure, Budd-Chiari syndrome, Byler disease, Crigler-Najjar syndrome, Caroli disease, cholestatic pruritus, cholelithiasis, conjugated hyperbilirubinemia, chronic graft-versus-host disease, cryptogenic liver disease, diabetes, Dubin-Johnson syndrome, erythrohepatic protoporphyria, extrahepatic bile duct carcinoma, familial hypercholesterolemia, galactosemia, Gilbert syndrome, glycogen storage disease, hemangioma, hemochromatosis, hepatic encephalopathy, hepatocholangitis, hepatomalacia, hepatomegalia, hepatocarcinoma, hepatoblastoma, hereditary hemochromatosis, jaundice, intrahepatic cholestasis, liver cysts, liver transplantation, liver failure associated with Bacillus cereus, mixed cryoglobulinemia, ornithine transcarbamylase deficiency, peliosis hepatis, porphyria cutanea tarda, primary biliary cirrhosis, refractory

ascites, Rotor syndrome, sarcoidosis, sclerosing cholangitis, steatosis, Summerskill syndrome, thrombocytopenia, tyrosinemia, variceal bleeding, venocclusive disease of the liver, and Wilson disease among many others, and are advantageously treated with the methods and compositions of the instant invention.

SUMM [0041] As a further embodiment of this invention a pharmaceutical composition is provided which is useful for treating and preventing a liver disease. The composition comprises an effective amount of cadaveric liver progenitor cells and a pharmaceutical carrier. The liver diseases of interest include acute or chronic liver disease of toxic, metabolic, genetic, and/or infective origin or of degenerative nature, or liver damage resulting from the use of drugs or substances injurious to the liver. Preferably among these conditions and diseases are inflammation of the liver, viral hepatitis, toxic liver cell damage, fibrosis of the liver, cirrhosis of the liver, liver congestion, liver dystrophy, fatty degeneration of liver cells, fatty liver, disturbances of the detoxification function, disturbances of the excretory function of the liver, disturbances of the conjugational function of the liver, disturbances of the synthesizing function of the liver portal hypertension due to a liver disease, or a liver failure coma, and intoxication by protein degradation products of ammonia. More specifically these include but are not limited to Alagille syndrome, alcoholic liver disease, alpha-1-antitrypsin deficiency, autoimmune hepatitis, biliary ductopenia, bone marrow failure, Budd-Chiari syndrome, biliary atresia, Byler disease, Crigler-Najjar syndrome, Caroli disease, cholestatic pruritus, cholelithiasis, conjugated hyperbilirubinemia, chronic graft-versus-host disease, cryptogenic liver disease, diabetes, Dubin-Johnson syndrome, erythrohepatic protoporphyria, extrahepatic bile duct carcinoma, familial hypercholesterolemia, galactosemia, Gilbert syndrome, glycogen storage disease, hemangioma, hemochromatosis, hepatic encephalopathy, hepatocholangitis, hepatomalacia, hepatomegalia, hepatocarcinoma, hepatoblastoma, hereditary hemochromatosis, jaundice, intrahepatic cholestasis, liver cysts, liver transplantation, liver failure associated with *Bacillus cereus*, mixed cryoglobulinemia, ornithine transcarbamylase deficiency, peliosis hepatis, porphyria cutanea tarda, primary biliary cirrhosis, refractory ascites, Rotor syndrome, sarcoidosis, sclerosing cholangitis, steatosis, Summerskill syndrome, thrombocytopenia, tyrosinemia, variceal bleeding, venocclusive disease of the liver, Wilson disease and combinations thereof.

DETD [0087] FIGS. 8a, 8b, 8c, 8d, 8e and 8f illustrate FACS analysis of fetal liver cell suspension for co-expression of CD14, CD38 and AFP. The bivariate scattergram (8a) shows the distribution of TriColor staining for CD14 (ordinate) versus FITC staining for CD38 (abscissa). Gates were created to select specific cell groupings according to the CD14 and CD38 signals. These were then used to display the intensity of AFP staining in each of these subgroups (FIGS. 8b, 8c, 8d and 8e). The AFP results show that a high level of enrichment for AFP is produced by selecting cells positive for either CD38 or CD14. The AFP signal generated from the entire cell suspension (30,000 cells) is shown in FIG. 8f.

DETD [0117] In most cases, the presence of AFP in the subgroups selected by cell surface marker is distributed continuously with a clear preponderance of cells showing staining intensities in the positive range. However, the distribution of CD38 positive cells with respect to co-expression of AFP is unique. In CD38-positive cells a bimodal distribution for AFP co-expression is apparent in which two distinct groups of cells are apparent, one group positive for AFP, the other



negative. This is illustrated in FIG. 8a which shows a scattergram of cells stained for expression of CD14 and CD38 together with univariate histograms of alpha-fetoprotein expression in cells positive for CD14 and/or CD 38.

DETD [0288] These and other useful applications are obvious to those skilled in the art. The specific examples of foreseen liver diseases include but are not limited to Alagille syndrome, alcoholic liver disease, alpha-1-antitrypsin deficiency, autoimmune hepatitis, biliary atresia, biliary ductopenia, bone marrow failure, Budd-Chiari syndrome, Byler disease, Crigler-Najjar syndrome, Caroli disease, cholestatic pruritus, cholelithiasis, conjugated hyperbilirubinemia, chronic graft-versus-host disease, cryptogenic liver disease, diabetes, Dubin-Johnson syndrome, erythrohepatic protoporphyria, extrahepatic bile duct carcinoma, familial hypercholesterolemia, galactosemia, Gilbert syndrome, glycogen storage disease, hemangioma, hemochromatosis, hepatic encephalopathy, hepatocholangitis, hepatomalacia, hepatomegalia, hepatocarcinoma, hepatoblastoma, hereditary hemochromatosis, jaundice, intrahepatic cholestasis, liver cysts, liver transplantation, liver failure associated with *Bacillus cereus*, mixed cryoglobulinemia, omithine transcarbamylase deficiency, peliosis hepatis, porphyria cutanea tarda, primary biliary cirrhosis, refractory ascites, Rotor syndrome, sarcoidosis, sclerosing cholangitis, steatosis, Summerskill syndrome, thrombocytopenia, tyrosinemia, variceal bleeding, venoocclusive disease of the liver, and Wilson disease.

L9 ANSWER 31 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2002:276193 USPATFULL <<LOGINID::20090129>>  
 TITLE: Cloning and characterization of genes encoding bradykinin B1 receptor homologues from five mammalian species  
 INVENTOR(S): Horlick, Robert, Cambridge, MA, United States  
 Zhao, Jiuqiao, Hockessin, DE, United States  
 Swanson, Robert, Cranbury, NJ, United States  
 Webb, Maria, Flemington, NJ, United States  
 Strohl, Barbara, Hamilton, NJ, United States  
 PATENT ASSIGNEE(S): Pharmacoepia, Inc., Cranbury, NJ, United States (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 6469150	B1	20021022
APPLICATION INFO.:	US 2000-576160		20000522 (9)
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	GRANTED		
PRIMARY EXAMINER:	Arthur, Lisa B.		
ASSISTANT EXAMINER:	Goldberg, Jeanine		
LEGAL REPRESENTATIVE:	Heslin Rothenberg Farley & Mesiti P.C.		
NUMBER OF CLAIMS:	8		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	15 Drawing Figure(s); 15 Drawing Page(s)		
LINE COUNT:	1308		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD Representative examples of appropriate hosts include bacterial cells, such as *E. coli*, *Streptomyces* and *Bacillus subtilis* cells; fungal cells, such as yeast cells and *Aspergillus* cells; insect cells such as *Drosophila* S2 and *Spodoptera* Sf9 cells; animal cells and plant cells. One of skill in the art will recognize that different host cells have characteristic and specific mechanisms for the post-translational processing and modification of proteins and gene products. Host cells suitable for expression of the inserted bradykinin receptor sequences of the present invention are those having the capability to effect such

post-translational modifications as necessary to produce a functional bradykinin receptor. Suitable mammalian host cells include but are not limited to CHO, VERO, BHK, HeLa, COS, MDCK, HEK 293, 3T3, W138.

DETD A subset of compounds that retain significant potency at most of the receptors was identified. Comparison of the activity of the compounds at animal vs. human B.sub.1 receptor orthologues is shown in the scattergrams of FIGS. 4a-b. The solid line at 45° in each panel represents an isocline of equal potency. FIGS. 4a-b show two independent human data sets compared to each other, and species-to-human comparisons, as labeled at the top of each scattergram. The correlation coefficients of each pair of data sets is shown below each figure. Compounds were tested at 1 µM concentration at the rat B.sub.1 receptor, and at 0.1 µM concentration at all other animal orthologues. Displacement was tested in the presence of 1.5 nM [<sup>3</sup>H]-dAKd for rat B.sub.1, and 0.6 nM for all other B.sub.1 receptors. Data points are marked as follow: .quadrature., PS978163; .diamond., PS596668; .largecircle., PS972282; Δ, PS309799. Conversely, a subset of compounds that exhibit considerable differences in specificity among the orthologues was also identified (data point for PS309799 shown enclosed by triangle, FIG. 4b). To verify the validity of the scattergram results, the potencies of these four non-peptide compounds were further assessed by ligand displacement assays at the B.sub.1 orthologous receptors. A comparison of K.sub.IS among the four compounds revealed dramatic differences in species specificity. Compound PS309799 showed the greatest variation of activity, ranging from low nM potency in tree shrew and human to inactive at dog and rabbit. PS596668 had a similar activity profile to PS309799 except it demonstrated potent activity at the rabbit B1. The remaining two compounds, PS972282 and PS978163, had measurable affinity constants at all six species, although PS978163 was considerably weaker at pig and dog.

L9 ANSWER 32 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2002:275894 USPATFULL <<LOGINID::20090129>>  
TITLE: Electrochemiluminescent rhenium moieties  
INVENTOR(S): Massey, Richard J., Rockville, MD, United States  
Powell, Michael J., Gaithersburg, MD, United States  
Dressick, Walter J., Gaithersburg, MD, United States  
Leland, Jonathan K., Gaithersburg, MD, United States  
Hino, Janel K., Arlington, VA, United States  
Poonian, Mohindar S., Gaithersburg, MD, United States  
Ciana, Leopoldo Della, Gaithersburg, MD, United States  
PATENT ASSIGNEE(S): IGEN International, Inc., Gaithersburg, MD, United States (U.S. corporation)

	NUMBER	KIND	DATE
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PATENT INFORMATION:	US 6468741	B1	20021022
APPLICATION INFO.:	US 1998-157788		19980921 (9)
RELATED APPLN. INFO.:	Continuation of Ser. No. US 1995-468524, filed on 6 Jun 1995, now patented, Pat. No. US 5811236 Division of Ser. No. US 1994-227898, filed on 15 Apr 1994, now patented, Pat. No. US 5591581 Continuation of Ser. No. US 1990-533931, filed on 5 Jun 1990, now abandoned Continuation of Ser. No. US 1987-117017, filed on 4 Nov 1987, now abandoned Continuation of Ser. No. US 1986-858354, filed on 30 Apr 1986, now abandoned		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	GRANTED		
PRIMARY EXAMINER:	Marschel, Ardin H.		
LEGAL REPRESENTATIVE:	Kramer Levin Naftalis & Frankel LLP, Evans, Esq., Barry		

NUMBER OF CLAIMS: 16  
EXEMPLARY CLAIM: 1  
NUMBER OF DRAWINGS: 3 Drawing Figure(s); 3 Drawing Page(s)  
LINE COUNT: 2864

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD The analytes of interest may be microorganisms. The microorganisms may be viable or nonviable or may be bacteria. Examples of bacteria which may be detected by this method include, but are not limited to, Salmonella, Campylobacter, Escherichia, Yersinia, Bacillus, Vibrio, Legionella, Clostridium, Streptococcus or Staphylococcus.

DETD Each serum sample is also analyzed for the concentration of theophylline by a fluorescence polarization assay. The concentration of theophylline measured by the homogeneous electrochemiluminescence immunoassay and the fluorescence polarization assay are compared. The data are plotted as a scattergram. The data points are analyzed by linear regression and the correlation coefficients are calculated. The analysis demonstrates an excellent correlation between the two assays. The correlation coefficients (r) are high. The slopes of the curves for normal, hemolyzed, and lipemic serum samples are between 0.8 and 1.2, demonstrating excellent recovery of theophylline from these serum samples.

L9 ANSWER 33 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2001:202782 USPATFULL <<LOGINID::20090129>>

TITLE: Electrochemiluminescent assays

INVENTOR(S): Massey, Richard J., Rockville, MD, United States  
Powell, Michael J., Rockville, MD, United States  
Mied, Paul A., New Windsor, MD, United States  
Feng, Peter, Rockville, MD, United States  
Della Ciana, Leopoldo, Rockville, MD, United States  
Dressick, Walter J., Rockville, MD, United States  
Poonian, Mohindar S., Gaithersburg, MD, United States  
PATENT ASSIGNEE(S): IGEN International, Inc., Gaithersburg, MD, United States (U.S. corporation)

	NUMBER	KIND	DATE
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PATENT INFORMATION:	US 6316607	B1	20011113
APPLICATION INFO.:	US 1995-472425		19950607 (8)
RELATED APPLN. INFO.:	Division of Ser. No. US 1995-415756, filed on 3 Apr 1995, now abandoned Continuation of Ser. No. US 1994-195825, filed on 10 Feb 1994, now abandoned Continuation of Ser. No. US 369560, now abandoned Continuation-in-part of Ser. No. US 1986-858354, filed on 30 Apr 1986, now abandoned		

DOCUMENT TYPE: Utility

FILE SEGMENT: GRANTED

PRIMARY EXAMINER: Riley, Jezia

LEGAL REPRESENTATIVE: Kramer Levin Naftalis & Frankel LLP

NUMBER OF CLAIMS: 46

EXEMPLARY CLAIM: 1

NUMBER OF DRAWINGS: 13 Drawing Figure(s); 13 Drawing Page(s)

LINE COUNT: 4227

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD The analytes of interest may be microorganisms. The microorganisms may be viable or nonviable. Additionally, the microorganisms may be bacteria. Examples of bacteria which may be detected by this method include, but are not limited to, Salmonella, Campylobacter, Escherichia, Yersinia, Bacillus, Vibrio, Legionella, Clostridium, Streptococcus or Staphylococcus.

DETD Each serum sample was also analyzed for the concentration of

theophylline by a fluorescence polarization assay. The concentration of theophylline measured by the homogeneous electrochemiluminescence immunoassay and the fluorescence polarization assay were compared. The data were plotted as a scattergram and are shown in FIGS.

4A-D. The data points were analyzed by linear regression and the correlation coefficients were calculated. The analysis demonstrates an excellent correlation between the two assays. The correlation coefficients (r) were between 0.98 and 1.00. The slopes of the curves for normal, hemolyzed, and lipemic serum samples were between 0.8 and 1.2, demonstrating excellent recovery of theophylline from these serum samples.

DETD The results for the homogeneous electrochemiluminescent immunoassay and the HPLC assay for determining the concentration of theophylline in serum are shown in FIG. 5. The data were plotted as a scattergram and the data points were analyzed by linear regression. The correlation coefficient was calculated. The correlation coefficient (r) was 0.98, which demonstrates excellent correlation between the two assays.

L9 ANSWER 34 OF 42 USPATFULL on STN

ACCESSION NUMBER: 2000:7062 USPATFULL <<LOGINID::20090129>>

TITLE: Antibody recognizing endothelial cell ligand for leukocyte CR3

INVENTOR(S): Tuomanen, Elaine, New York, NY, United States

Masure, H. Robert, New York, NY, United States

PATENT ASSIGNEE(S): The Rockefeller University, New York, NY, United States (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 6015560		20000118
APPLICATION INFO.:	US 1995-465966		19950606 (8)
RELATED APPLN. INFO.:	Division of Ser. No. US 1994-348353, filed on 30 Nov 1994 which is a continuation-in-part of Ser. No. US 1994-247572, filed on 23 May 1994, now abandoned which is a continuation of Ser. No. WO 1992-US3725, filed on 4 May 1992 which is a continuation-in-part of Ser. No. US 1991-695613, filed on 3 May 1991, now abandoned		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	Granted		
PRIMARY EXAMINER:	Minnifield, Nita		
LEGAL REPRESENTATIVE:	Klauber & Jackson		
NUMBER OF CLAIMS:	14		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	31 Drawing Figure(s); 42 Drawing Page(s)		
LINE COUNT:	3341		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM The process of this invention will be useful in treating inflammation caused by any of a variety of infective agents, including gram-positive and gram-negative bacteria as well as viruses and fungi. Particularly targeted infections are those which are susceptible to treatment with beta-lactam antibiotics, or antiviral agents such as Haemophilus influenzae B; N. meningitidis b; pneumococci, e.g., Streptococcus pneumoniae; Escherichia coli; Staphylococcus epidermidus; Staphylococcus aureus; group B Streptococci; Salmonella; Bacillus subtilis; Pseudomonas aeruginosa; and Herpes virus.

DRWD FIG. 22 is a scattergram representation of the effect of variation of the FHA peptide II structure on efficacy of inhibition of meningeal inflammation. Values are leukocyte densities at 7 hours for individual rabbits. The horizontal lines indicate the mean and standard

deviation of CSF leukocyte density in 10 control animals which received phosphate buffered saline.

DRWD FIG. 23 is a scattergram representation of the ability of acetyl/amide FHA peptide II to inhibit accumulation of leukocytes in the CSF. Two groups of 10 animals were challenged with pneumococci. One hour later, the animals received an intravenous injection of 10 nmoles of the acetyl/amide peptide (8.2 µg) or phosphate buffered saline. Leukocyte density in CSF was determined 6 hours after pneumococcal challenge. The mean values as indicated by the bars are statistically significantly different at p=0.0015 by ANOVA.

L9 ANSWER 35 OF 42 USPATFULL on STN

ACCESSION NUMBER: 1999:128131 USPATFULL <<LOGINID::20090129>>

TITLE: Antibody recognizing endothelial cell ligand for leukocyte CR3

INVENTOR(S): Tuomanen, Elaine, New York, NY, United States  
Masure, H. Robert, New York, NY, United States

PATENT ASSIGNEE(S): The Rockefeller University, New York, NY, United States  
(U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 5968512		19991019
APPLICATION INFO.:	US 1995-465965		19950606 (8)
RELATED APPLN. INFO.:	Division of Ser. No. US 1994-348353, filed on 30 Nov 1994 which is a continuation-in-part of Ser. No. US 1994-247572, filed on 23 May 1994, now abandoned which is a continuation of Ser. No. WO 1992-US3725, filed on 4 May 1992 which is a continuation-in-part of Ser. No. US 1991-695613, filed on 3 May 1991, now abandoned		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	Granted		
PRIMARY EXAMINER:	Minnifield, Nita		
LEGAL REPRESENTATIVE:	Klauber & Jackson		
NUMBER OF CLAIMS:	3		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	47 Drawing Figure(s); 42 Drawing Page(s)		
LINE COUNT:	3297		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM The process of this invention will be useful in treating inflammation caused by any of a variety of infective agents, including gram-positive and gram-negative bacteria as well as viruses and fungi. Particularly targeted infections are those which are susceptible to treatment with beta-lactam antibiotics, or antiviral agents such as Haemophilus influenzae B; N. meningitidis b; pneumococci, e.g., Streptococcus pneumoniae; Escherichia coli; Staphylococcus epidermidus; Staphylococcus aureus; group B Streptococci; Salmonella; Bacillus subtilis; Pseudomonas aeruginosa; and Herpes virus.

DRWD FIG. 22 is a scattergram representation of the effect of variation of the FHA peptide II structure on efficacy of inhibition of meningeal inflammation. Values are leukocyte densities at 7 hours for individual rabbits. The horizontal lines indicate the mean and standard deviation of CSF leukocyte density in 10 control animals which received phosphate buffered saline.

DRWD FIG. 23 is a scattergram representation of the ability of acetyl/amide FHA peptide II to inhibit accumulation of leukocytes in the CSF. Two groups of 10 animals were challenged with pneumococci. One hour later, the animals received an intravenous injection of 10 nmoles of the acetyl/amide peptide (8.2 µg) or phosphate buffered saline. Leukocyte density in CSF was determined 6 hours after pneumococcal challenge. The

mean values as indicated by the bars are statistically significantly different at p=0.0015 by ANOVA.

L9 ANSWER 36 OF 42 USPATFULL on STN

ACCESSION NUMBER: 1999:88796 USPATFULL <<LOGINID::20090129>>  
TITLE: Peptides which inhibit adhesion between leukocytes and endothelial cells  
INVENTOR(S): Tuomanen, Elaine, New York, NY, United States  
Masure, H. Robert, New York, NY, United States  
PATENT ASSIGNEE(S): The Rockefeller University, New York, NY, United States (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 5932217		19990803
APPLICATION INFO.:	US 1994-348353		19941130 (8)
RELATED APPLN. INFO.:	Continuation-in-part of Ser. No. US 1994-247572, filed on 23 May 1994, now abandoned which is a continuation-in-part of Ser. No. US 140136		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	Granted		
PRIMARY EXAMINER:	Caputa, Anthony C.		
ASSISTANT EXAMINER:	Navarro, Mark		
LEGAL REPRESENTATIVE:	Klauber & Jackson		
NUMBER OF CLAIMS:	8		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	37 Drawing Figure(s); 42 Drawing Page(s)		
LINE COUNT:	3167		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM The process of this invention will be useful in treating inflammation caused by any of a variety of infective agents, including gram-positive and gram-negative bacteria as well as viruses and fungi. Particularly targeted infections are those which are susceptible to treatment with beta-lactam antibiotics, or antiviral agents such as Haemophilus influenzae B; N. meningitidis b; pneumococci, e.g., Streptococcus pneumoniae; Escherichia coli; Staphylococcus epidermidis; Staphylococcus aureus; group B Streptococci; Salmonella; Bacillus subtilis; Pseudomonas aeruginosa; and Herpes virus.

DRWD FIG. 22 is a scattergram representation of the effect of variation of the FHA peptide II structure on efficacy of inhibition of meningeal inflammation. Values are leukocyte densities at 7 hours for individual rabbits. The horizontal lines indicate the mean and standard deviation of CSF leukocyte density in 10 control animals which received phosphate buffered saline.

DRWD FIG. 23 is a scattergram representation of the ability of acetyl/amide FHA peptide II to inhibit accumulation of leukocytes in the CSF. Two groups of 10 animals were challenged with pneumococci. One hour later, the animals received an intravenous injection of 10 nmoles of the acetyl/amide peptide (8.2 µg) or phosphate buffered saline. Leukocyte density in CSF was determined 6 hours after pneumococcal challenge. The mean values as indicated by the bars are statistically significantly different at p=0.0015 by ANOVA.

L9 ANSWER 37 OF 42 USPATFULL on STN

ACCESSION NUMBER: 1998:115553 USPATFULL <<LOGINID::20090129>>  
TITLE: Electrochemiluminescent rhenium moieties and methods for their use  
INVENTOR(S): Massey, Richard J., Rockville, MD, United States  
Powell, Michael J., Gaithersburg, MD, United States  
Dressick, Walter J., Gaithersburg, MD, United States

## PATENT ASSIGNEE(S):

Leland, Jonathan K., Gaithersburg, MD, United States  
Hino, Janel K., Arlington, VA, United States  
Poonian, Mohindar S., Gaithersburg, MD, United States  
Ciana, Leopoldo Della, Gaithersburg, MD, United States  
IGEN International, Inc., Gaithersburg, MD, United States (U.S. corporation)

	NUMBER	KIND	DATE
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PATENT INFORMATION:	US 5811236		19980922
APPLICATION INFO.:	US 1995-468524		19950606 (8)
RELATED APPLN. INFO.:	Division of Ser. No. 227898, filed on 15 Apr 1994 which is a continuation of Ser. No. 533931, filed on 5 Jun 1990, now abandoned which is a continuation of Ser. No. 117017, filed on 4 Nov 1987, now abandoned which is a continuation-in-part of Ser. No. 858354, filed on 30 Apr 1986, now abandoned		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	Granted		
PRIMARY EXAMINER:	Jones, W. Gary		
ASSISTANT EXAMINER:	Rees, Dianne		
LEGAL REPRESENTATIVE:	Curtis, Morris & Safford, P.C., Evans, Barry, Rubin, David		
NUMBER OF CLAIMS:	131		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	3 Drawing Figure(s); 3 Drawing Page(s)		
LINE COUNT:	3565		

## CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD The analytes of interest may be microorganisms. The microorganisms may be viable or nonviable or may be bacteria. Examples of bacteria which may be detected by this method include, but are not limited to, Salmonella, Campylobacter, Escherichia, Yersinia, Bacillus, Vibrio, Legionella, Clostridium, Streptococcus or Staphylococcus.

DETD Each serum sample is also analyzed for the concentration of theophylline by a fluorescence polarization assay. The concentration of theophylline measured by the homogeneous electrochemiluminescence immunoassay and the fluorescence polarization assay are compared. The data are plotted as a scattergram. The data points are analyzed by linear regression and the correlation coefficients are calculated. The analysis demonstrates an excellent correlation between the two assays. The correlation coefficients (r) are high. The slopes of the curves for normal, hemolyzed, and lipemic serum samples are between 0.8 and 1.2, demonstrating excellent recovery of theophylline from these serum samples.

L9 ANSWER 38 OF 42 USPATFULL on STN

ACCESSION NUMBER: 1998:95235 USPATFULL <<LOGINID::20090129>>

TITLE: Antibody recognizing endothelial cell ligand for leukocyte CR3

INVENTOR(S): Tuomanen, Elaine, New York, NY, United States  
Masure, H. Robert, New York, NY, United States

PATENT ASSIGNEE(S): The Rockefeller University, New York, NY, United States (U.S. corporation)

	NUMBER	KIND	DATE
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PATENT INFORMATION:	US 5792457		19980811
APPLICATION INFO.:	US 1995-465929		19950606 (8)
RELATED APPLN. INFO.:	Division of Ser. No. US 1994-348353, filed on 30 Nov 1994 which is a continuation-in-part of Ser. No. US 1994-247572, filed on 23 May 1994, now abandoned which		

is a continuation-in-part of Ser. No. US 1991-695613,  
filed on 3 May 1991, now abandoned

DOCUMENT TYPE: Utility  
FILE SEGMENT: Granted  
PRIMARY EXAMINER: Hutzell, Paula K.  
ASSISTANT EXAMINER: Krikorian, Jacqueline G.  
LEGAL REPRESENTATIVE: Klauber & Jackson  
NUMBER OF CLAIMS: 9  
EXEMPLARY CLAIM: 1  
NUMBER OF DRAWINGS: 47 Drawing Figure(s); 41 Drawing Page(s)  
LINE COUNT: 2578  
CAS INDEXING IS AVAILABLE FOR THIS PATENT.

SUMM The process of this invention will be useful in treating inflammation caused by any of a variety of infective agents, including gram-positive and gram-negative bacteria as well as viruses and fungi. Particularly targeted infections are those which are susceptible to treatment with beta-lactam antibiotics, or antiviral agents such as Haemophilus influenzae B; N. meningitidis b; pneumococci, e.g., Streptococcus pneumoniae; Escherichia coli; Staphylococcus epidermidis; Staphylococcus aureus; group B Streptococci; Salmonella; Bacillus subtilis; Pseudomonas aeruginosa; and Herpes virus.

DRWD FIG. 22 is a scattergram representation of the effect of variation of the FHA peptide II structure on efficacy of inhibition, of meningeal inflammation. Values are leukocyte densities at 7 hours for individual rabbits. The horizontal lines indicate the mean and standard deviation of CSF leukocyte density in 10 control animals which received phosphate buffered saline.

DRWD FIG. 23 is a scattergram representation of the ability of acetyl/amide FHA peptide II to inhibit accumulation of leukocytes in the CSF. Two groups of 10 animals were challenged with pneumococci. One hour later, the animals received an intravenous injection of 10 nmoles of the acetyl/amide peptide (8.2 µg) or phosphate buffered saline. Leukocyte density in CSF was determined 6 hours after pneumococcal challenge. The mean values as indicated by the bars are statistically significantly different at p=0.0015 by ANOVA.

L9 ANSWER 39 OF 42 USPATFULL on STN

ACCESSION NUMBER: 1998:14635 USPATFULL <<LOGINID::20090129>>  
TITLE: Method of calibration of an electrochemiluminescent assay system

INVENTOR(S): Massey, Richard J., Rockville, MD, United States  
Powell, Michael J., Gaithersburg, MD, United States  
Dressick, Walter J., Gaithersburg, MD, United States  
Leland, Jonathan K., Gaithersburg, MD, United States  
Hino, Janel K., Arlington, VA, United States  
Poonian, Mohindar S., Gaithersburg, MD, United States  
Ciana, Leopoldo Della, Gaithersburg, MD, United States  
PATENT ASSIGNEE(S): Igen International Inc., Gaithersburg, MD, United States (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 5716781		19980210
APPLICATION INFO.:	US 1995-470247		19950606 (8)
RELATED APPLN. INFO.:	Division of Ser. No. US 1994-227898, filed on 15 Apr 1994, now patented, Pat. No. US 5591581 which is a continuation of Ser. No. US 1990-533931, filed on 5 Jun 1990, now abandoned which is a continuation of Ser. No. US 1987-117017, filed on 4 Nov 1987, now abandoned which is a continuation-in-part of Ser. No. US		



1986-858354, filed on 30 Apr 1986, now abandoned

DOCUMENT TYPE: Utility  
 FILE SEGMENT: Granted  
 PRIMARY EXAMINER: Marschel, Ardin H.  
 ASSISTANT EXAMINER: Riley, Jezia  
 LEGAL REPRESENTATIVE: Curtis, Morris & Safford, P.C., Evans, Esq., Barry, Rubin, Esq., David

NUMBER OF CLAIMS: 12  
 EXEMPLARY CLAIM: 1  
 NUMBER OF DRAWINGS: 3 Drawing Figure(s); 3 Drawing Page(s)  
 LINE COUNT: 2978  
 CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD The analytes of interest may be microorganisms. The microorganisms may be viable or nonviable or may be bacteria. Examples of bacteria which may be detected by this method include, but are not limited to, Salmonella, Campylobacter, Escherichia, Yersinia, Bacillus, Vibrio, Legionella, Clostridium, Streptococcus or Staphylococcus.

DETD Each serum sample is also analyzed for the concentration of theophylline by a fluorescence polarization assay. The concentration of theophylline measured by the homogeneous electrochemiluminescence immunoassay and the fluorescence polarization assay are compared. The data are plotted as a scattergram. The data points are analyzed by linear regression and the correlation coefficients are calculated. The analysis demonstrates an excellent correlation between the two assays. The correlation coefficients (r) are high. The slopes of the curves for normal, hemolyzed, and lipemic serum samples are between 0.8 and 1.2, demonstrating excellent recovery of theophylline from these serum samples.

L9 ANSWER 40 OF 42 USPATFULL on STN

ACCESSION NUMBER: 97:1313 USPATFULL <<LOGINID::20090129>>  
 TITLE: Electrochemiluminescent rhenium moieties and methods for their use

INVENTOR(S): Massey, Richard J., Rockville, MD, United States  
 Powell, Michael J., Gaithersburg, MD, United States  
 Dressick, Walter J., Gaithersburg, MD, United States  
 Leland, Jonathan K., Gaithersburg, MD, United States  
 Hino, Janel K., Arlington, VA, United States  
 Poonian, Mohindar S., Gaithersburg, MD, United States  
 Ciana, Leopoldo D., Gaithersburg, MD, United States

PATENT ASSIGNEE(S): IGEN, Inc., Rockville, MD, United States (U.S. corporation)

	NUMBER	KIND	DATE
PATENT INFORMATION:	US 5591581		19970107
APPLICATION INFO.:	US 1994-227898		19940415 (8)
RELATED APPLN. INFO.:	Continuation of Ser. No. US 1990-533931, filed on 5 Jun 1990, now abandoned which is a continuation of Ser. No. US 1987-117017, filed on 4 Nov 1987, now abandoned which is a continuation-in-part of Ser. No. US 1986-858354, filed on 30 Apr 1986		
DOCUMENT TYPE:	Utility		
FILE SEGMENT:	Granted		
PRIMARY EXAMINER:	Nucker, Christine M.		
ASSISTANT EXAMINER:	Stucker, Jeffrey		
LEGAL REPRESENTATIVE:	Curtis Morris & Safford, P.C.		
NUMBER OF CLAIMS:	20		
EXEMPLARY CLAIM:	1		
NUMBER OF DRAWINGS:	3 Drawing Figure(s); 3 Drawing Page(s)		
LINE COUNT:	2937		

CAS INDEXING IS AVAILABLE FOR THIS PATENT.

DETD The analytes of interest may be microorganisms. The microorganisms may be viable or nonviable or may be bacteria. Examples of bacteria which may be detected by this method include, but are not limited to, *Salmonella*, *Campylobacter*, *Escherichia*, *Yersinia*, *Bacillus*, *Vibrio*, *Legionella*, *Clostridium*, *Streptococcus* or *Staphylococcus*.

DETD Each serum sample is also analyzed for the concentration of theophylline by a fluorescence polarization assay. The concentration of theophylline measured by the homogeneous electrochemiluminescence immunoassay and the fluorescence polarization assay are compared. The data are plotted as a scattergram. The data points are analyzed by linear regression and the correlation coefficients are calculated. The analysis demonstrates an excellent correlation between the two assays. The correlation coefficients (r) are high. The slopes of the curves for normal, hemolyzed, and lipemic serum samples are between 0.8 and 1.2, demonstrating excellent recovery of theophylline from these serum samples.

L9 ANSWER 41 OF 42 CAPLUS COPYRIGHT 2009 ACS on STN

ACCESSION NUMBER: 1994:4412 CAPLUS <<LOGINID::20090129>>

DOCUMENT NUMBER: 120:4412

ORIGINAL REFERENCE NO.: 120:1003a,1006a

TITLE: In vitro activity of Bay y 3118, a new quinolone

AUTHOR(S): Fass, Robert J.

CORPORATE SOURCE: Coll. Med., Ohio State Univ., Columbus, OH, 43210, USA

SOURCE: Antimicrobial Agents and Chemotherapy (1993), 37(11), 2348-57

CODEN: AMACCQ; ISSN: 0066-4804

DOCUMENT TYPE: Journal

LANGUAGE: English

AB MICs of Bay y 3118, ciprofloxacin, ofloxacin, clarithromycin, azithromycin, cefuroxime, amoxicillin-clavulanate, and trimethoprim-sulfamethoxazole for 878 recent clin. isolates were determined by broth microdiln. methods. Among the 3 quinolones, Bay y 3118 was the most active against *Haemophilus influenzae*, *Moraxella catarrhalis*, *Acinetobacter baumannii*, *Xanthomonas maltophilia*, gram-pos. cocci, and anaerobes; MICs for 50% of the strains (MIC50s) and MIC90s were  $\leq 0.015$  and  $\leq 0.015$ ,  $\leq 0.015$  and  $\leq 0.015$ , 0.03 and 2, 0.25 and 0.5, 0.06 and 1, and 0.12 and 0.25  $\mu\text{g/mL}$ , resp. For gram-pos. cocci and anaerobes, these values were 16- to 32-fold (4-5 log<sub>2</sub> dilution steps) lower than those for ciprofloxacin and ofloxacin. Bay y 3118 was similar in activity to ciprofloxacin and more active than ofloxacin against members of the family Enterobacteriaceae and *Pseudomonas aeruginosa*; Bay y 3118 MIC50s and MIC90s were 0.03 and 0.25 and 0.5 and 8  $\mu\text{g/mL}$ , resp. Scattergrams and regression analyses comparing quinolone MICs indicated that, despite differences in activity, organisms relatively susceptible to 1 were relatively susceptible to all and organisms relatively resistant to 1 were relatively resistant to all. However, the greater in vitro activity of Bay y 3118 was most pronounced against relatively resistant organisms. Pending pharmacokinetic and safety data for Bay y 3118, there is reasonable anticipation that its enhanced activity against gram-pos. cocci and anaerobes would broaden the clin. utility of the quinolone class of antimicrobial agents.

L9 ANSWER 42 OF 42 CAPLUS COPYRIGHT 2009 ACS on STN

ACCESSION NUMBER: 1994:3950 CAPLUS <<LOGINID::20090129>>

DOCUMENT NUMBER: 120:3950

ORIGINAL REFERENCE NO.: 120:911a,914a

TITLE: Erythromycin, clarithromycin, and azithromycin: use of frequency distribution curves, scattergrams, and regression analyses to compare in vitro activities

and describe cross-resistance

AUTHOR(S): Fass, R. J.

CORPORATE SOURCE: Coll. Med., Ohio State Univ., Columbus, OH, 43210, USA

SOURCE: Antimicrobial Agents and Chemotherapy (1993), 37(10), 2080-6

CODEN: AMACCQ; ISSN: 0066-4804

DOCUMENT TYPE: Journal

LANGUAGE: English

TI Erythromycin, clarithromycin, and azithromycin: use of frequency distribution curves, scattergrams, and regression analyses to compare in vitro activities and describe cross-resistance

AB MICs of erythromycin, clarithromycin, and azithromycin for 852 recent clin. isolates were determined by broth microdilution methods. Frequency distribution curves, scattergrams, and regression analyses were used to compare in vitro activities and describe cross-resistance. Clarithromycin was the most active drug against *Bacteroides* species but the least active against *Haemophilus influenzae*. Azithromycin was most active against *H. influenzae*, *Moraxella catarrhalis*, *Pasteurella multocida*, and *Fusobacterium* species but least active against *Streptococcus* species and *Enterococcus* species. All three drugs had equivalent activities against *Staphylococcus* species and gram.-pos. anaerobes. None of the three drugs was particularly active against members of the family Enterobacteriaceae or nonfermentative gram-neg. bacilli, although concns. of 4 µg/mL of azithromycin inhibited some strains of the family Enterobacteriaceae (particularly *Escherichia coli* and *Citrobacter diversus*) and *Acinetobacter baumannii*. Although relative drug activities varied by organism, organisms relatively susceptible to one were relatively susceptible to all and organisms relatively resistant to one were relatively resistant to all; an exception was fusobacteria, which were usually susceptible only to azithromycin. Cross-susceptibility and cross-resistance were, therefore, the rule (except for *Fusobacterium* species, although the percentage of susceptible organisms could be varied considerably on the basis of the selection of breakpoints).

IT *Acinetobacter baumannii*

*Bacteroides*

*Citrobacter diversus*

Enterobacteriaceae

*Escherichia coli*

*Fusobacterium*

*Haemophilus influenzae*

*Moraxella catarrhalis*

*Pasteurella multocida*

*Staphylococcus*

*Streptococcus*

(antibiotic sensitivity of, frequency distribution curves and scattergrams and regression analyses in study of)

IT Bacteria

(bacilli, gram-neg., antibiotic sensitivity of, frequency distribution curves and scattergrams and regression analyses in study of)

IT *Streptococcus*

(intestinal, antibiotic sensitivity of, frequency distribution curves and scattergrams and regression analyses in study of)

IT 114-07-8, Erythromycin 81103-11-9, Clarithromycin 83905-01-5, Azithromycin

RL: BIOL (Biological study)

(bacteria sensitivity to, frequency distribution curves and scattergrams and regression analyses in study of)